

Microarray hybridization: Learning about RNA- degradation, G-stacks and binding isotherms

© Hans Binder & Mario Fasold

University Leipzig

Interdisciplinary Centre for Bioinformatics



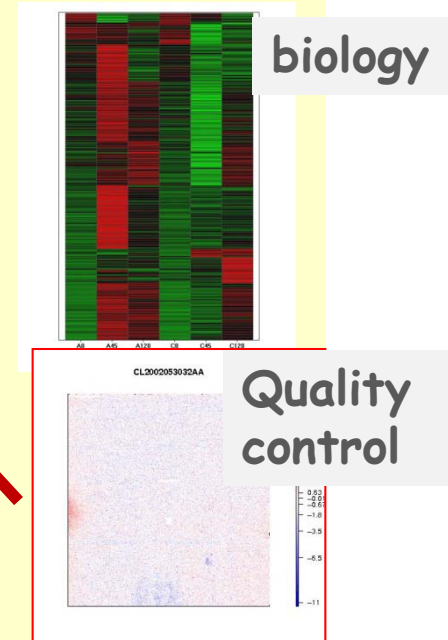
Plön May/2011

Microarrays are tools for studying

1. biology (expression, copy numbers, genotypes...)



Calibration:
extracting proper
expression values

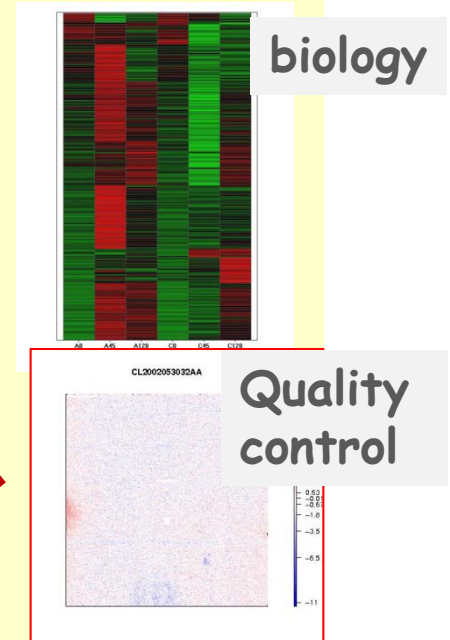


Microarrays are tools for studying

1. biology (expression, copy numbers, genotypes...)



Calibration:
extracting proper
expression values



2. surface hybridization (isotherms, molecular interactions)

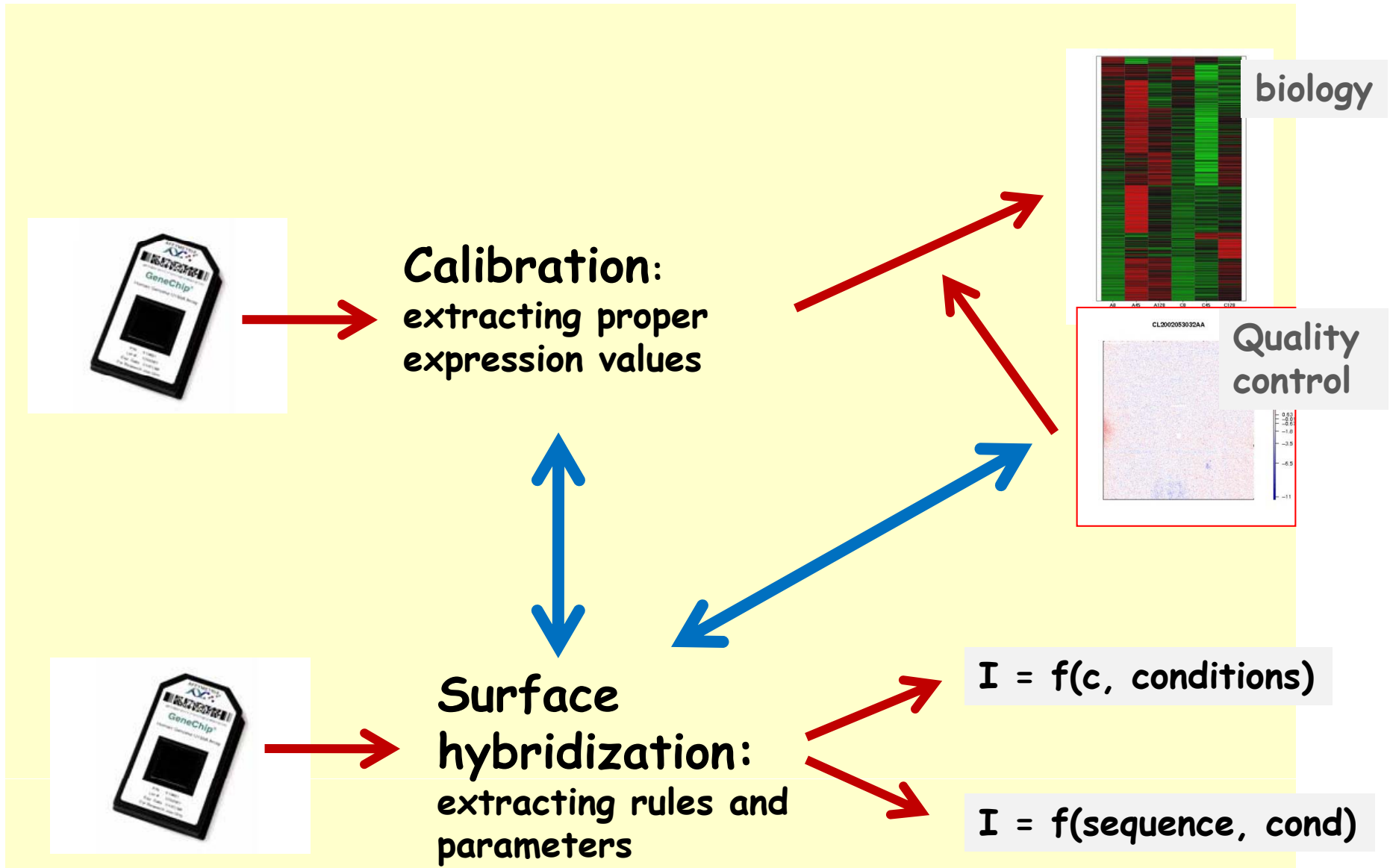


**Surface
hybridization:**
extracting rules and
parameters

$$I = f(c, \text{conditions})$$

$$I = f(\text{sequence}, \text{cond})$$

Microarrays are tools for studying



Bad and good things about microarrays

a) bad things about microarrays - effects which distort intensity

- non-specific background
- sequence-specific affinity
- saturation of probe spots
- RNA degradation, 3'/5'-bias
- specific sequence motifs: e.g. GGG



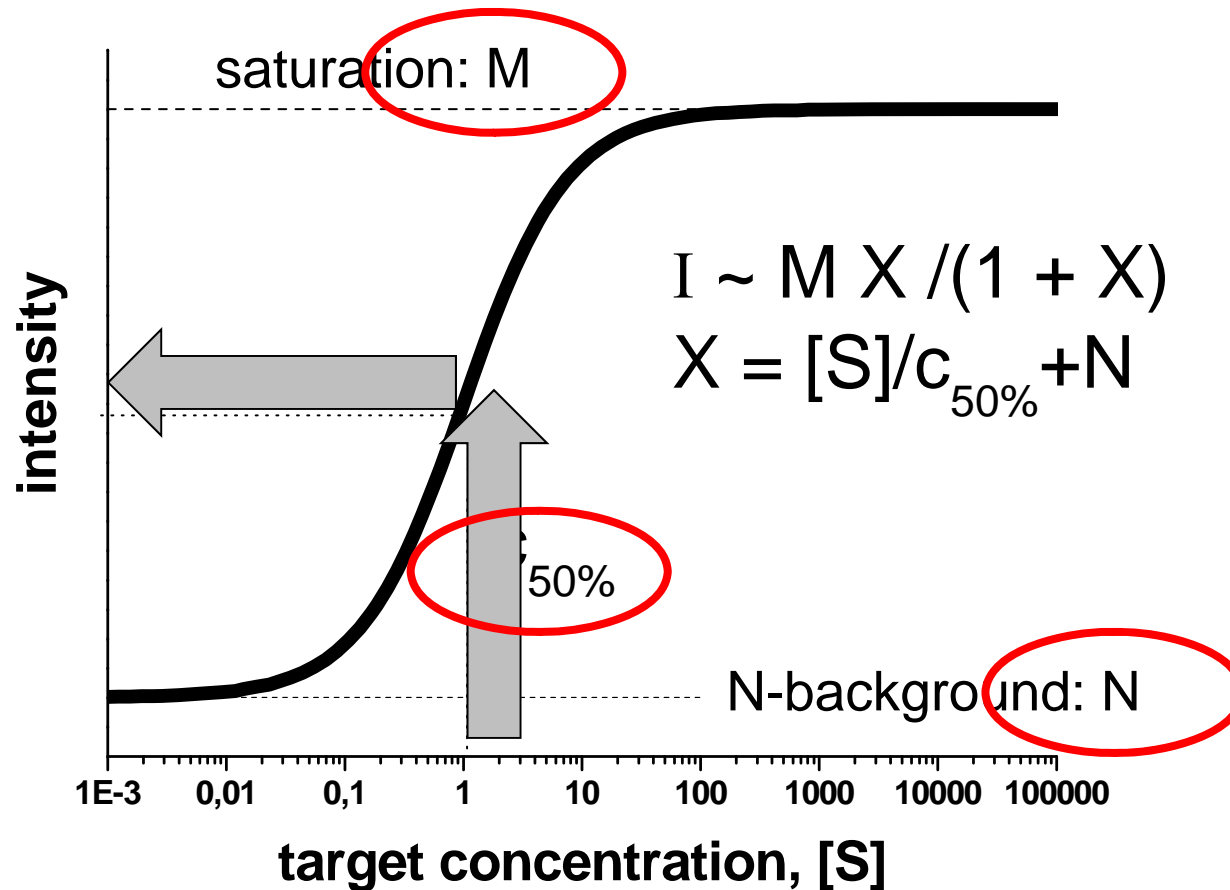
Good things:

- To solve: Suited rules (isotherms, interactions) and sufficient set-parameters that can be estimated for each array
- Physico-chemical stuff: e.g. degradation and GGG-effects

The inverse (Langmuir-) isotherm problem



Irvin Langmuir
(1881-1957)
Nobel prize 1932



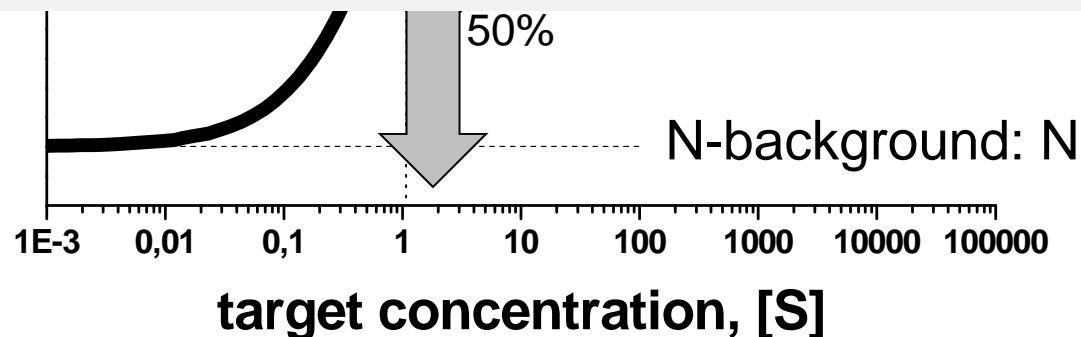
The inverse (Langmuir-) isotherm problem



saturation: M

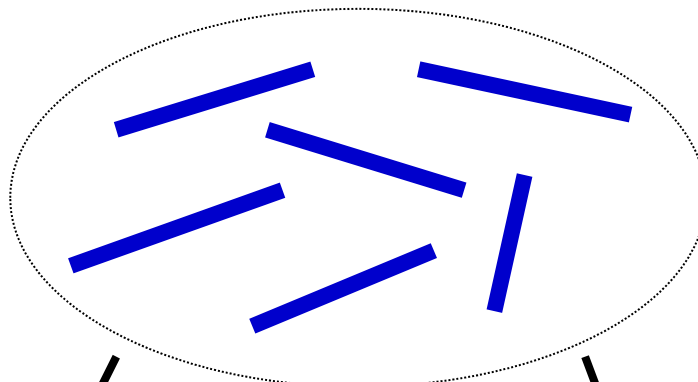
Options to estimate the parameters:

1. Special calibration data (e.g. spiked-in) for each array
2. Use 'ordinary' probe data as intrinsic reference ('hook' method)
 - Expression values (inverse Langmuir solution)
 - Quality control



Situation 1: PM/MM-pairings

targets



identical targets

strong affinity : $c_{50\%}^{probe}$

weak affinity : $c_{50\%}^{ref}$



PM

MM

,old affy' expression

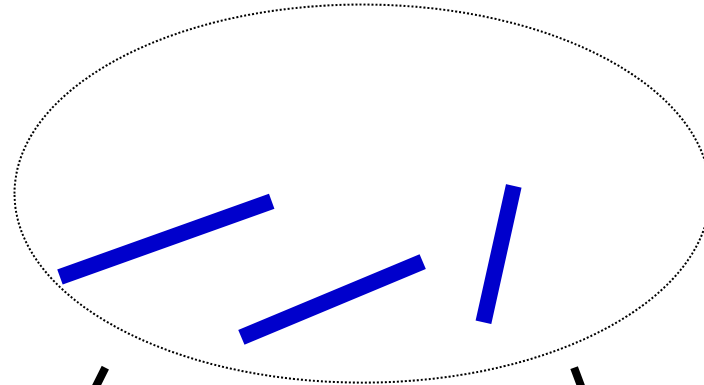
allele A

cross allele B

SNP-arrays

Situation 1: PM/MM-pairings

targets



strong affinity : $c_{50\%}^{probe}$

weak affinity : $c_{50\%}^{ref}$



probes

internal reference

microarray

PM

MM

,old affy' expression

allele A

allele B

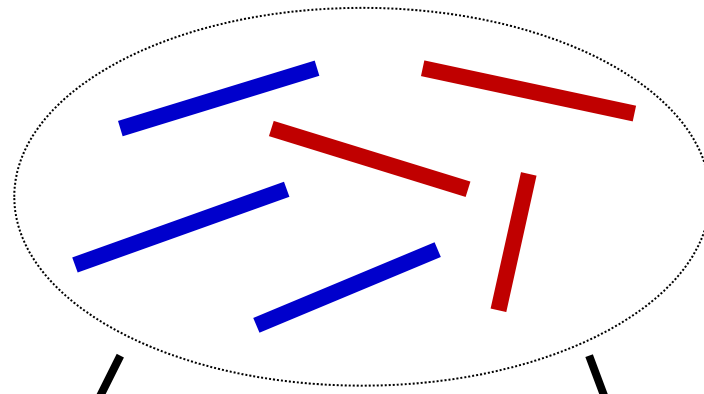
SNP-arrays

Matched

mismatched duplexes

Situation 2: PM-only

targets



Targets interrogating the same transcripts

strong affinity : $c_{50\%}^{probe}$

weak affinity : $c_{50\%}^{ref}$



PM(strong)

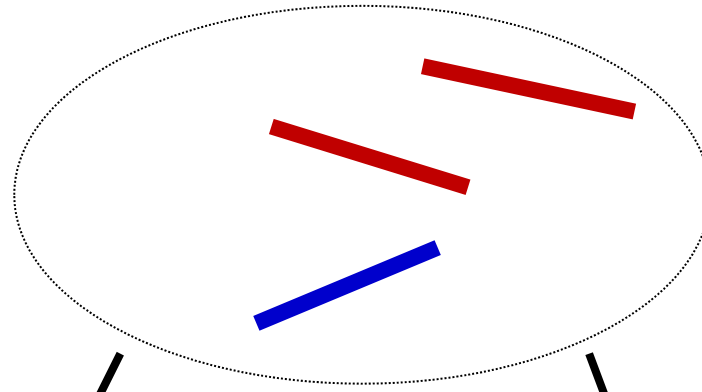
PM(weak)

PMonly expression

Strongly-bound weakly-bound matched duplexes

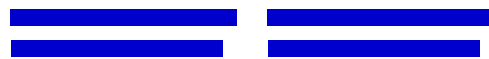
Situation 2: PM/PM-pairings

targets



strong affinity : $c_{50\%}^{\text{probe}}$

weak affinity : $c_{50\%}^{\text{ref}}$



probes

internal reference

microarray

PM(strong)

PM(weak)

PMonly expression

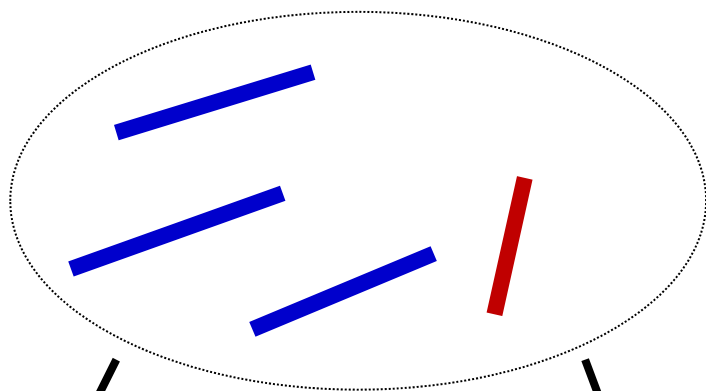
Strongly-bound

weakly-bound

matched duplexes

Situation 3: high/low-conc-pairings

targets



Targets interrogating the same transcripts

High conc.: $[S]^{high}$

Low conc.: $[S]^{low}$



PM(3')
PM(before wash)

PM(5')
PM(after wash)

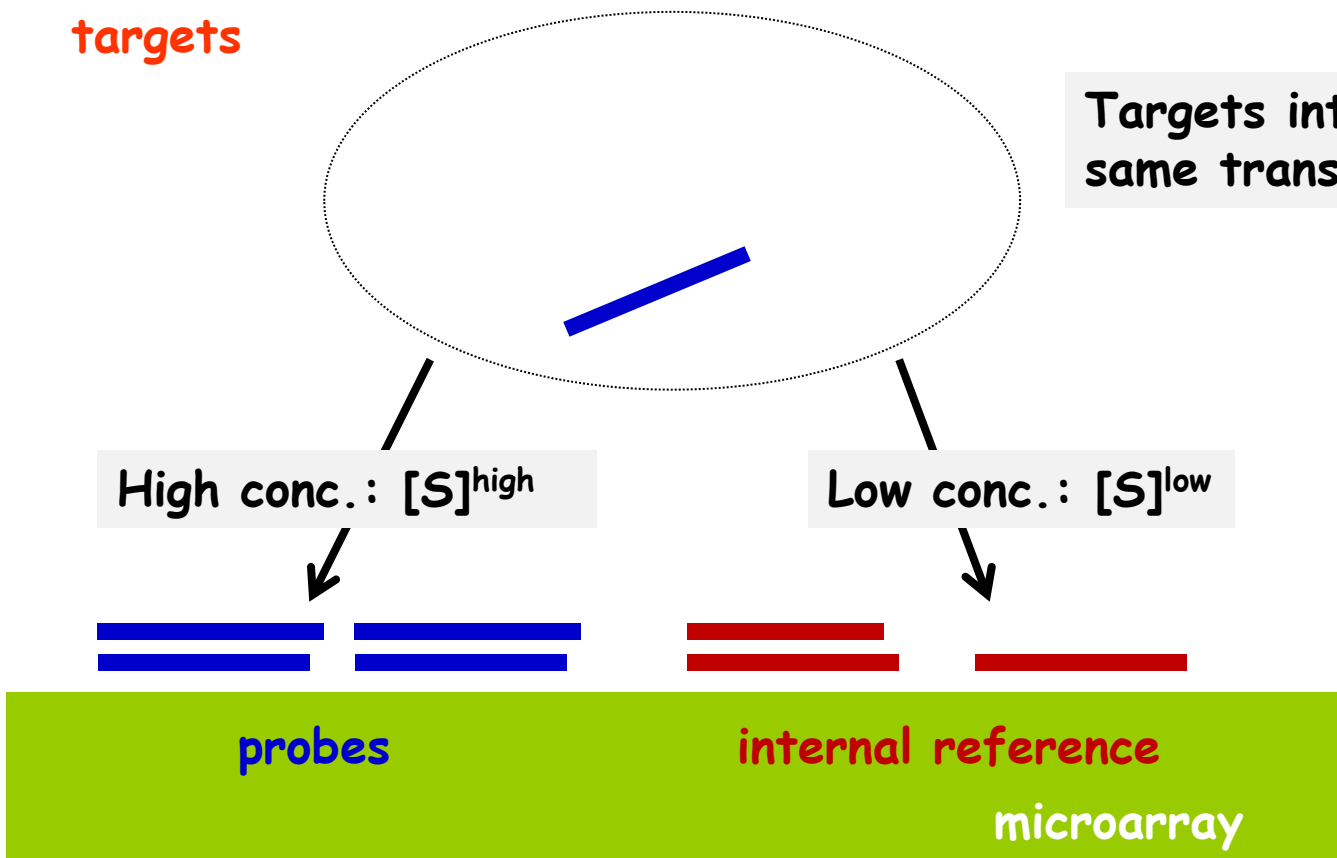
PMonly expression

Strongly-bound weakly-bound matched duplexes

Situation 3: high-conc/low-conc-pairings

targets

Targets interrogating the same transcripts



PM(3')
PM(before wash)

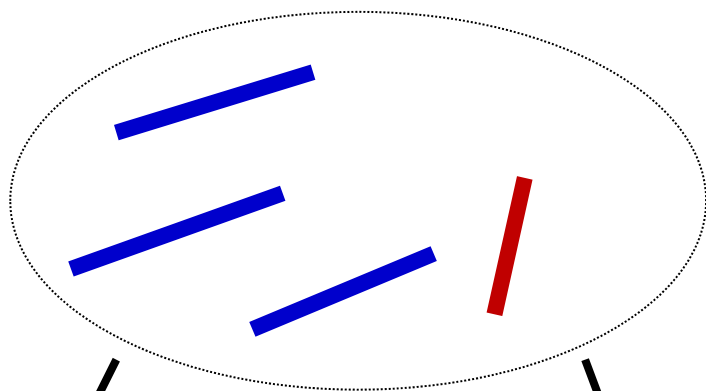
PM(5')
PM(after wash)

P-sets

Highly- weakly- populated duplexes

General: high/low-binding strength

targets



Targets interrogating the same transcripts

High : X

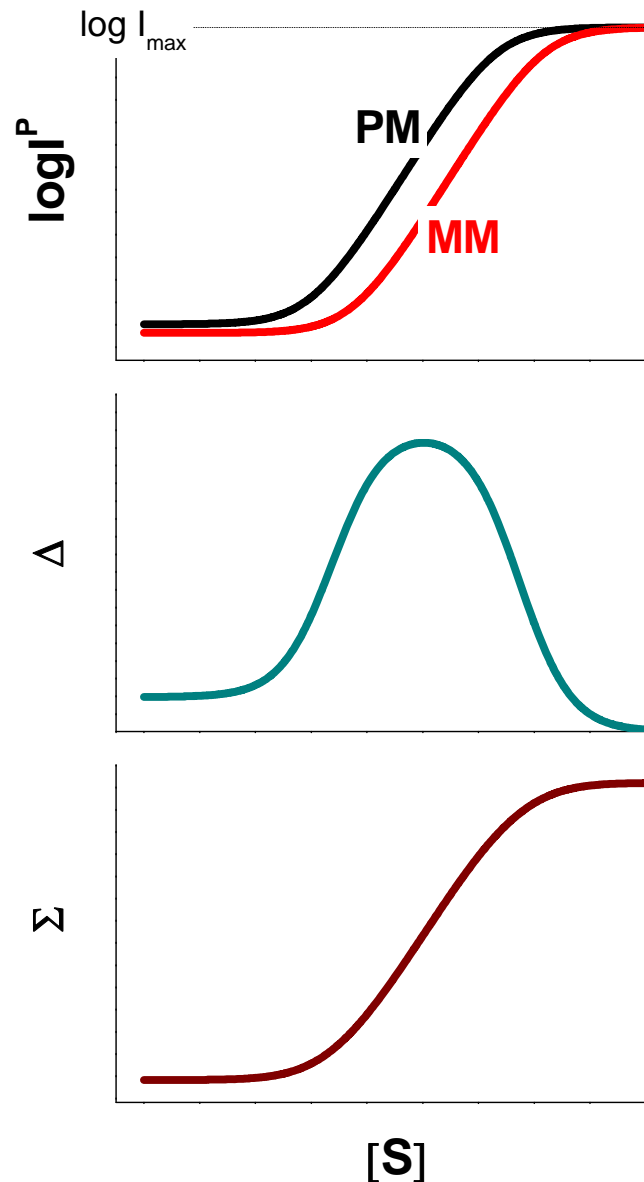
Low: X^{ref}

$$X = [S]/c_{50\%}$$



Highly- weakly- populated duplexes

Example: PM/MM-pairings



Let's use the MM as internal reference, by calculating

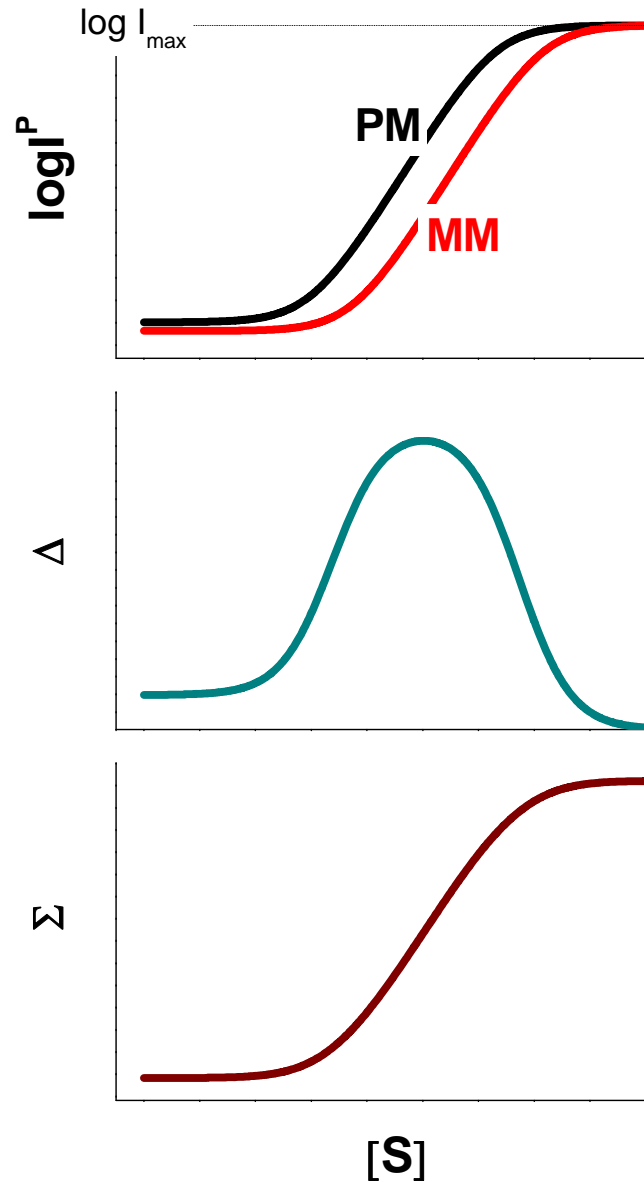
- the difference

$$\Delta \equiv \log PM - \log MM$$

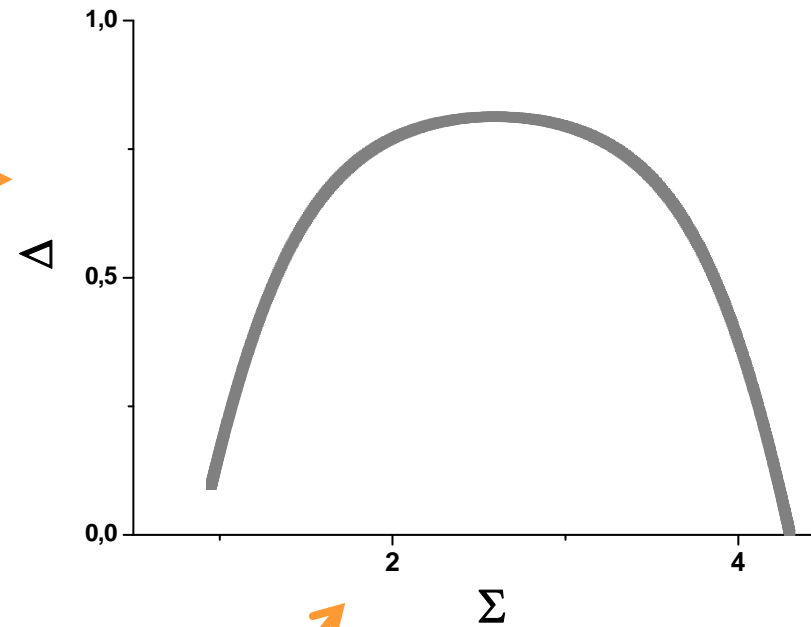
- the mean

$$\Sigma \equiv \frac{1}{2} (\log PM + \log MM)$$

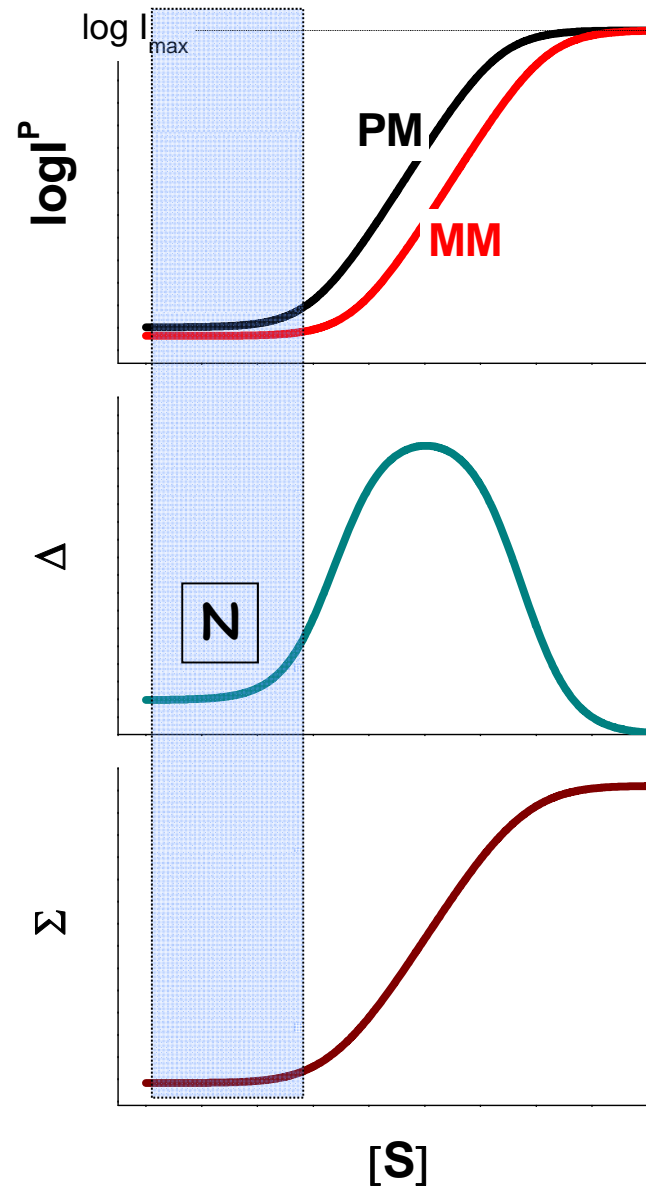
The Δ -vs- Σ plot



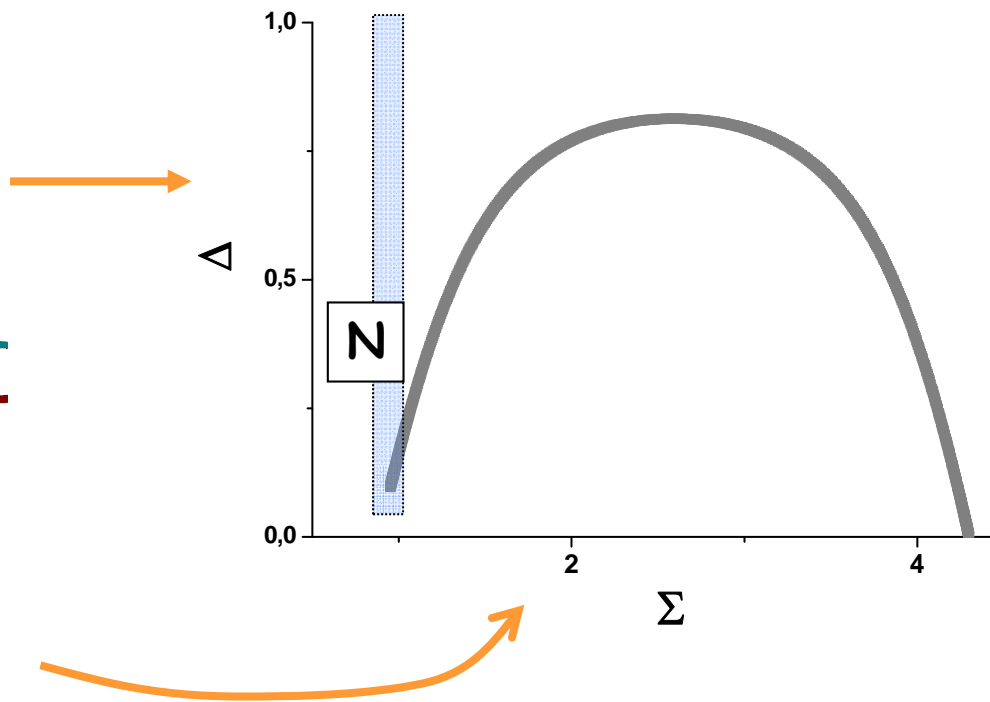
... and by plotting Δ -vs- Σ into a sort of M-A-plot



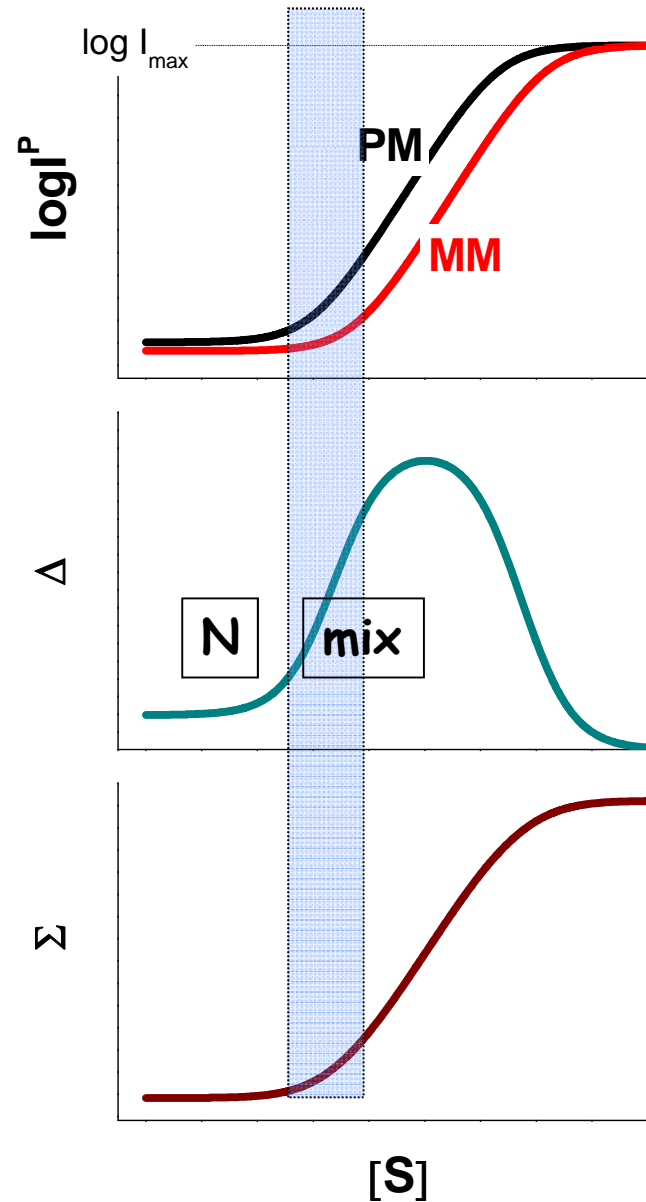
The hybridization regimes



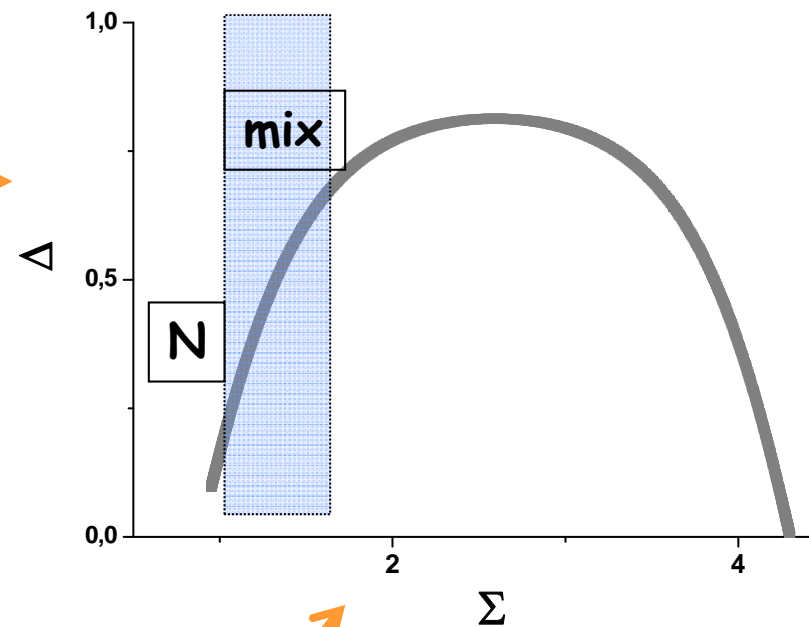
... can be clearly identified:
 N... non-specific binding



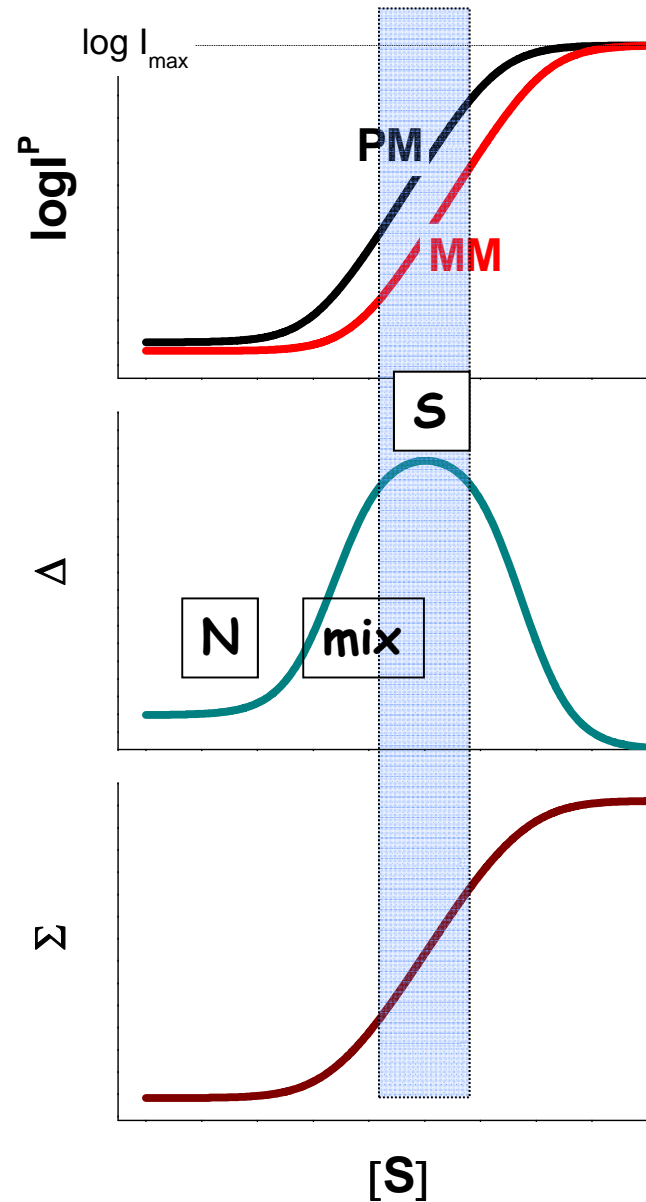
The hybridization regimes



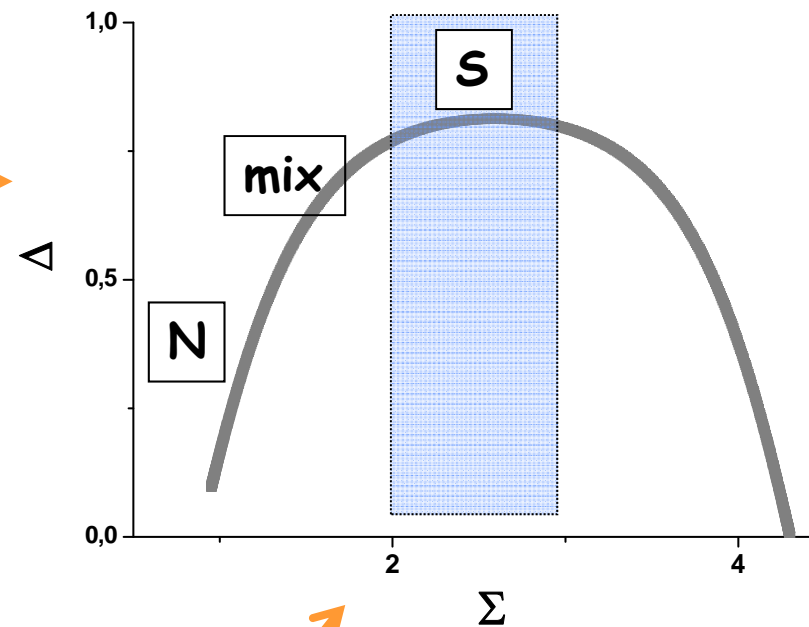
... can be clearly identified:
mix... N- and S-hybridization



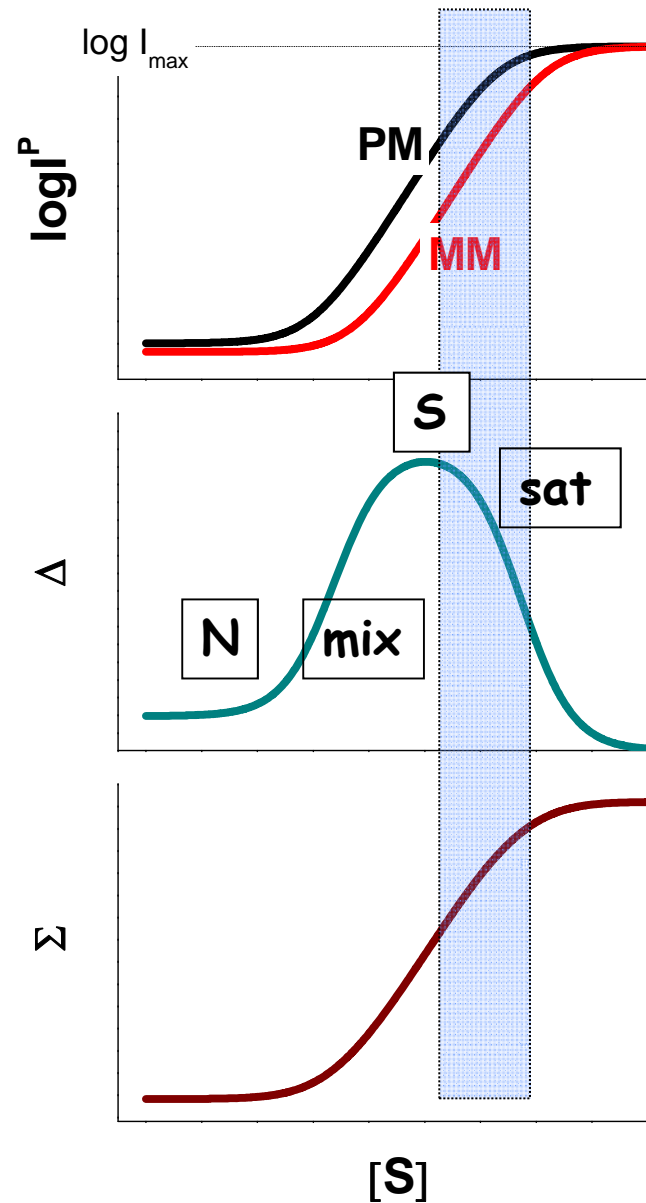
The hybridization regimes



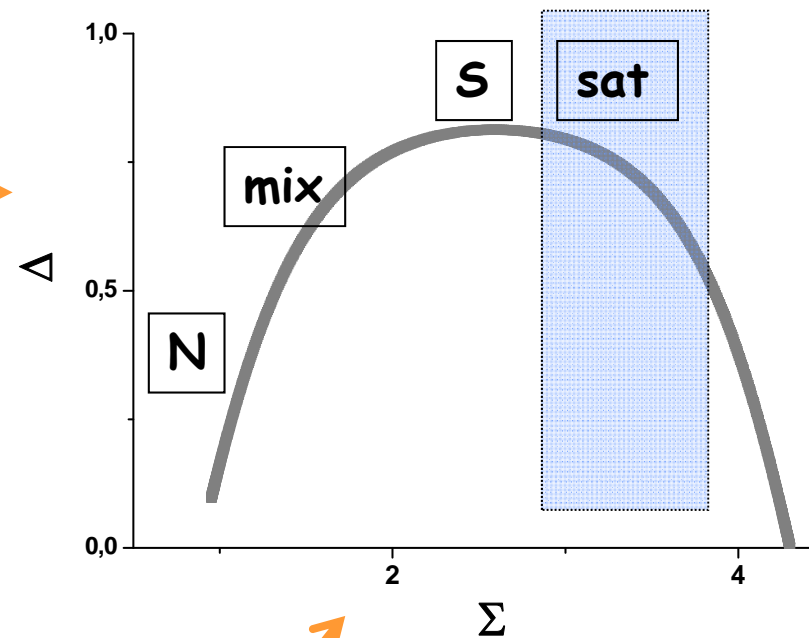
... can be clearly identified:
S... specific hybridization



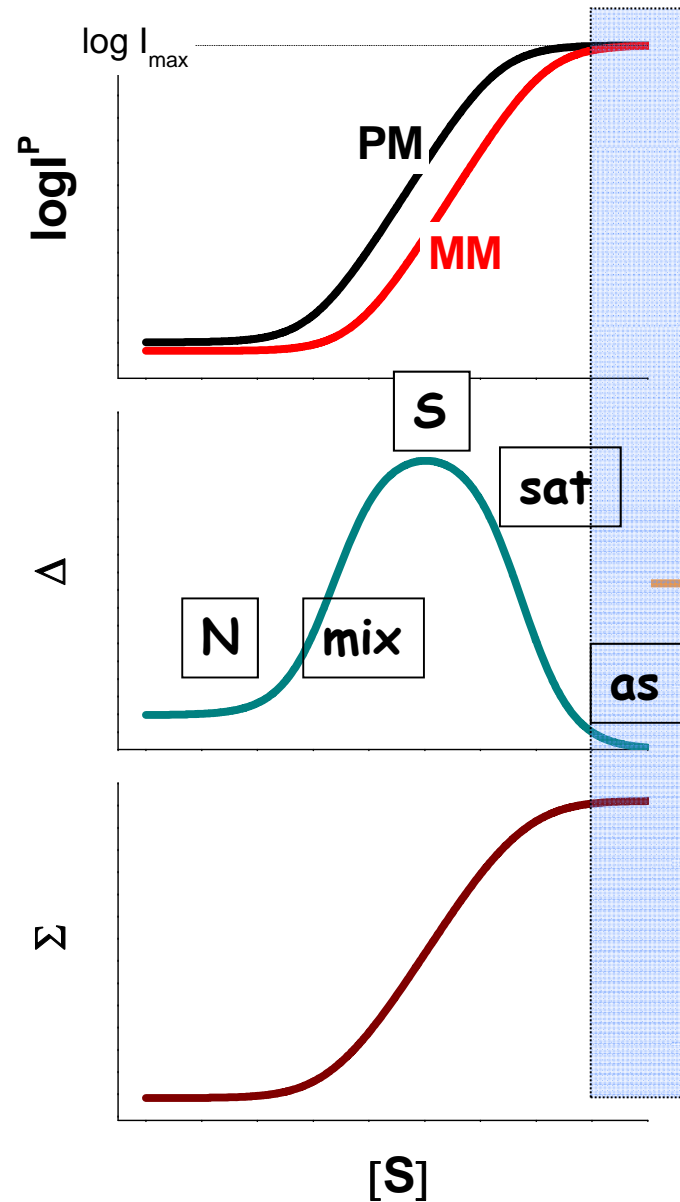
The hybridization regimes



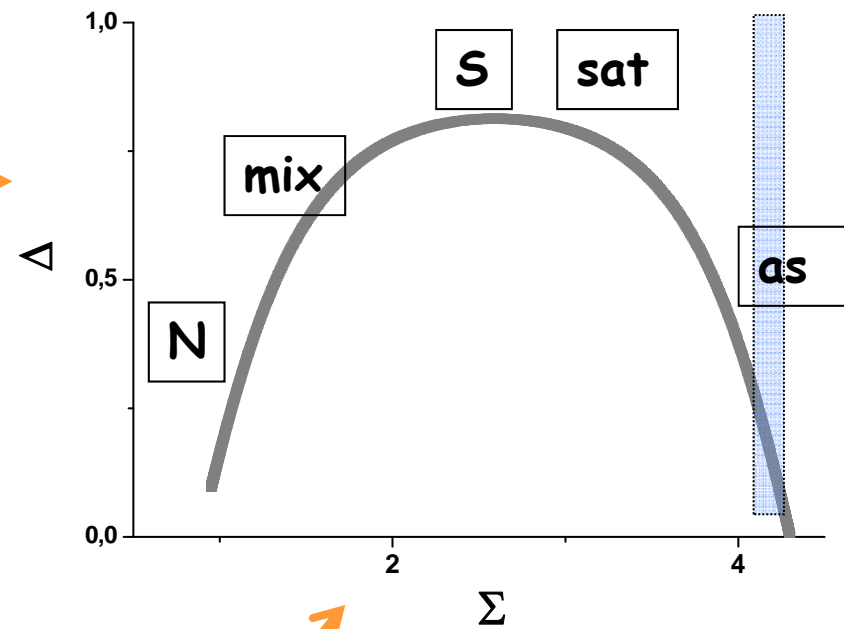
... can be clearly identified:
sat... saturation



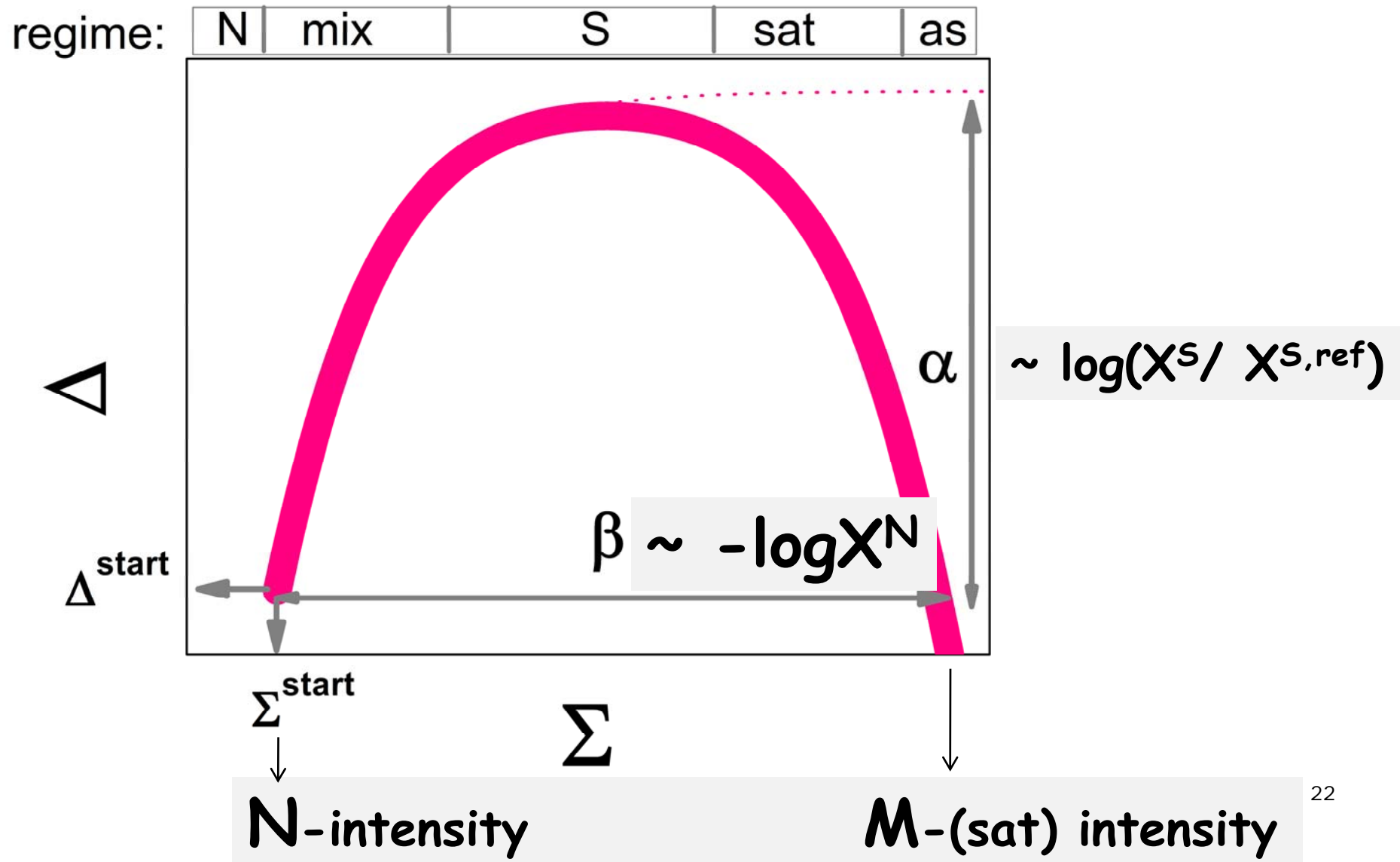
The hybridization regimes



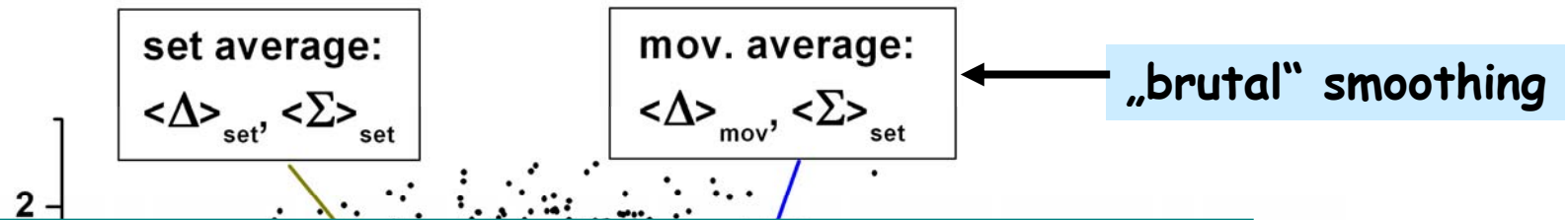
... can be clearly identified:
as... asymptotic



Position and dimensions of the hook curve

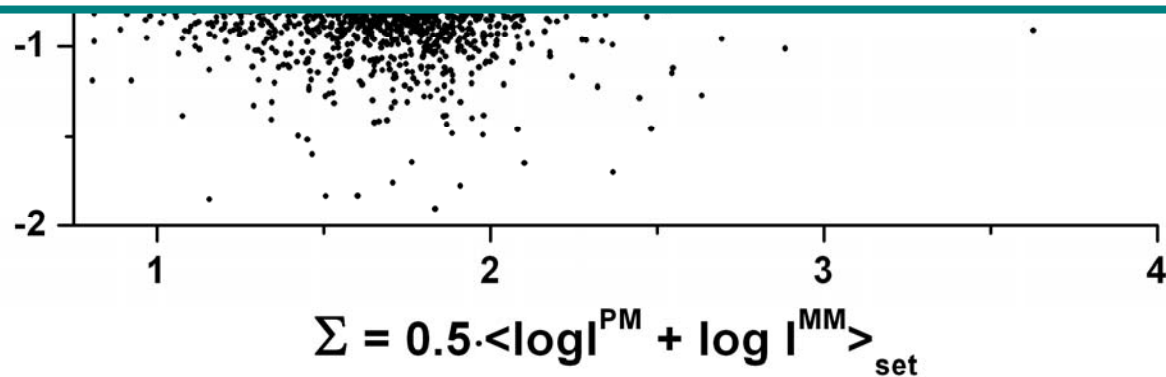


Mean Δ -vs- Σ plot of the chip

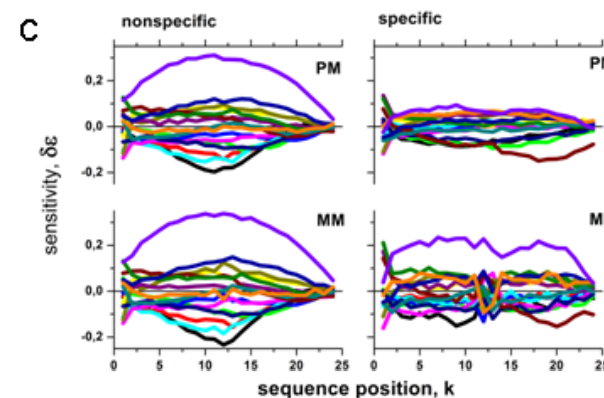
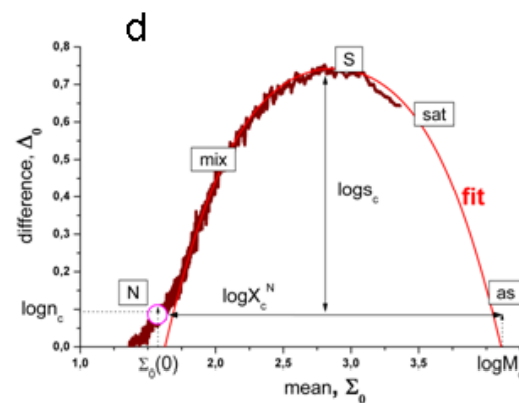
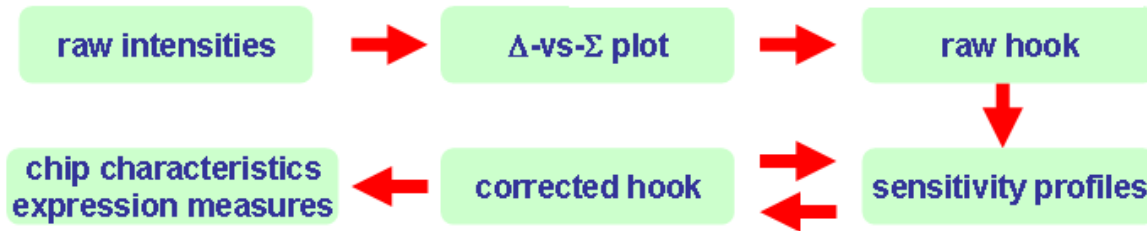
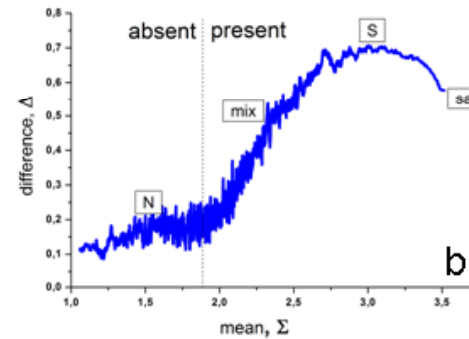
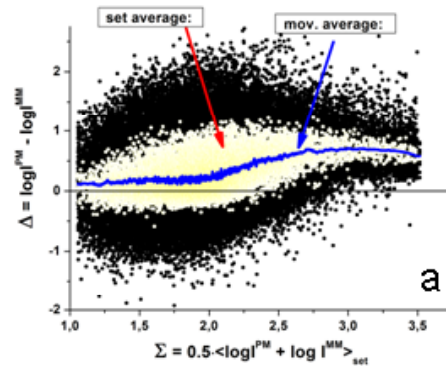


We consider all probe-pairs of a chip

- we calculate the average Δ -vs- Σ trajectory by applying heavy smoothing
- provides the mean isotherm of a particular hybridization

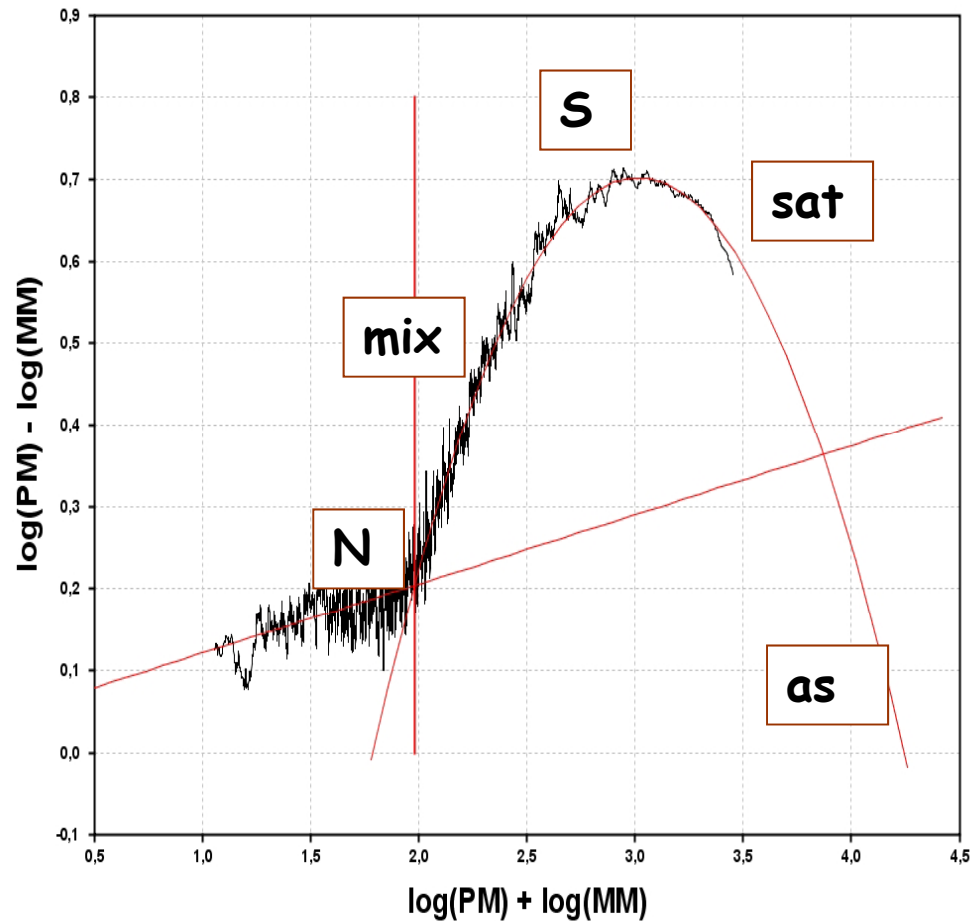


Hook analysis: overview



positional-dependent NN model

the (raw) hook - curve reflects the...



Hybridization regimes:

N...non-specific hybridization dominates

mix...N+S hybridization

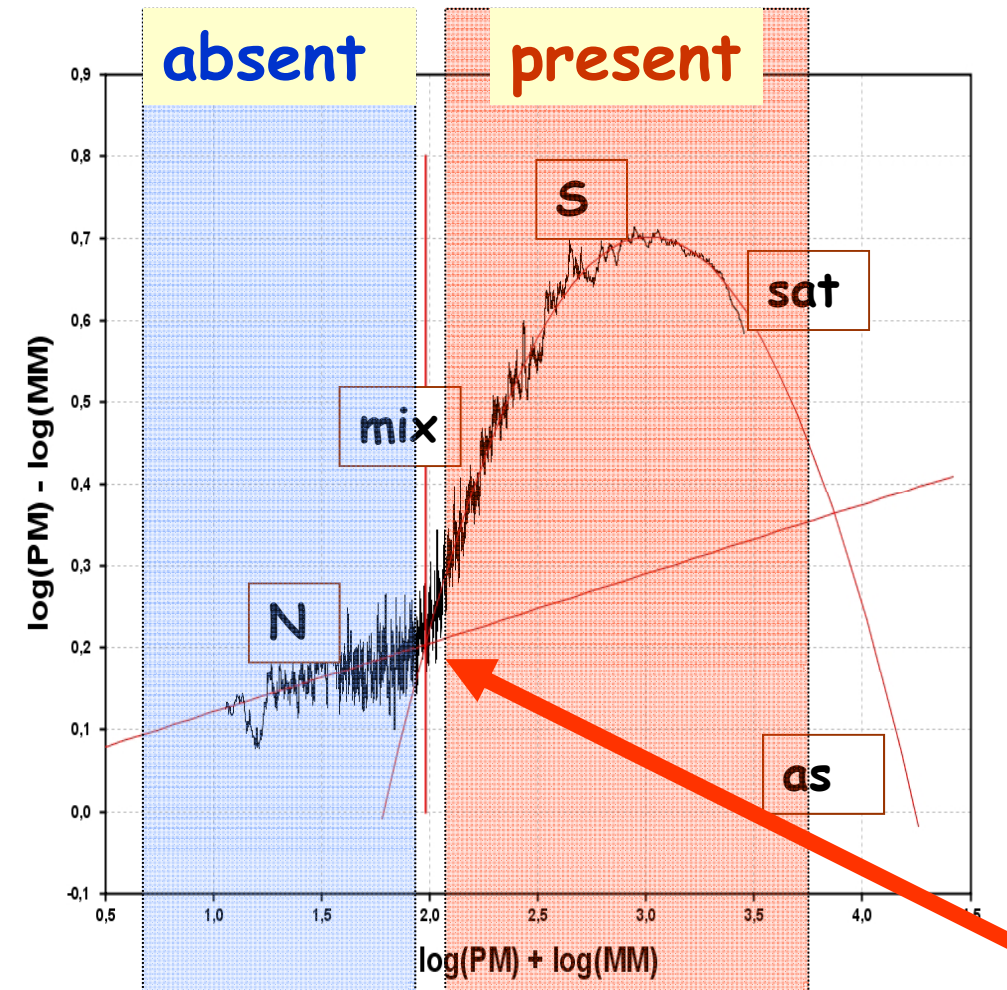
S... specific binding dominates

sat...progressive saturation

(as...asymptotic range, the probes are saturated)

...which we know from the single probes

Absent and present probes...



We suggest:
the initial break can be used
to identify „absent“ probes,

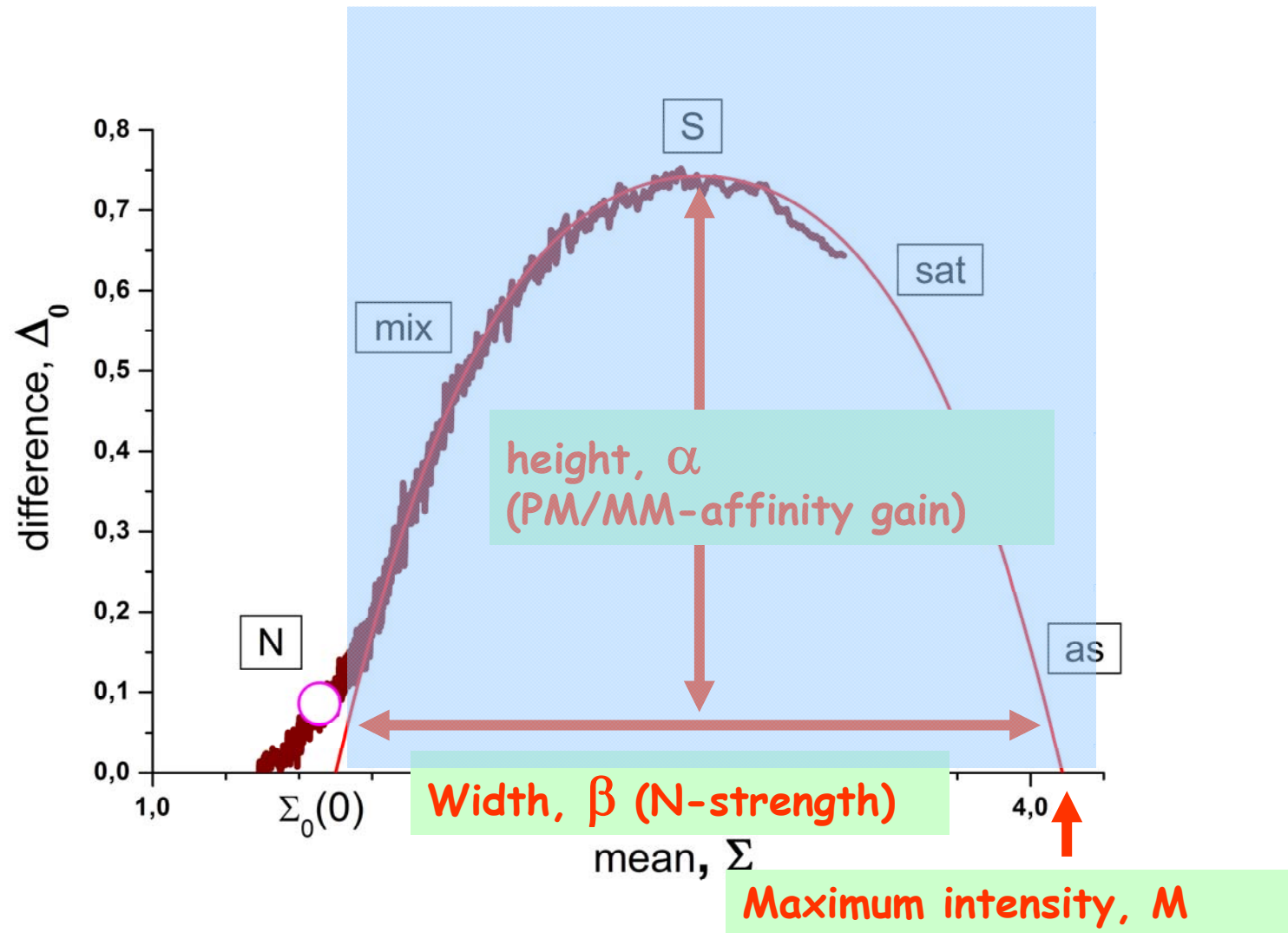
i.e. which predominantly
hybridize non-specifically
because their specific
transcripts are virtually absent
in the hybridization solution

slope(N) ~ correlations PM/MM

slope(mix) ~ S-dependence of
the isotherm

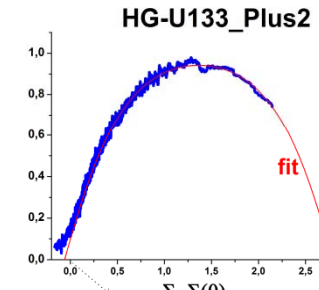
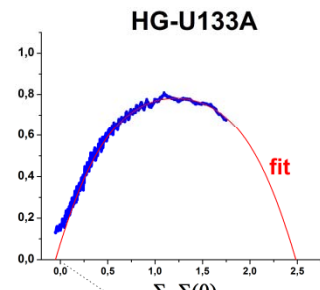
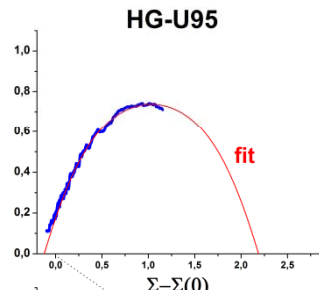
break

Fit of the hook-equation provides...

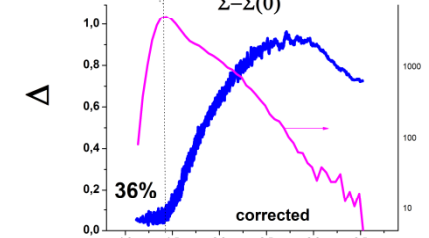
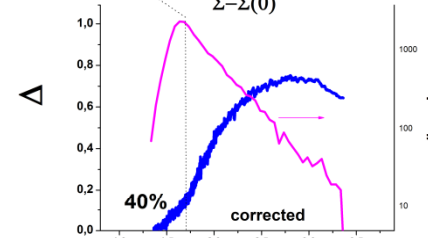
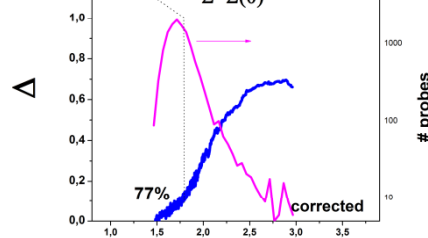


Chip generations (human genome)

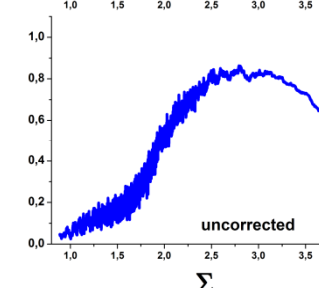
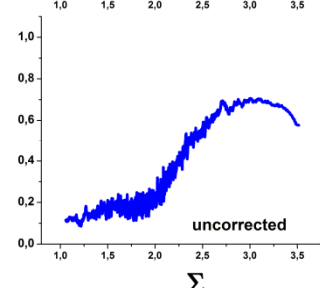
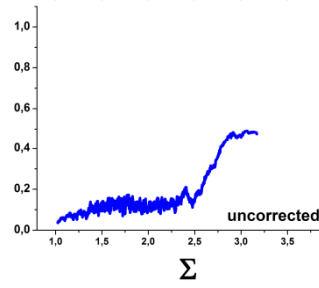
fit



cor-
rec-
ted



raw



Spot size: 20 μM

18 μM

11 μM

probe sets: 16,000

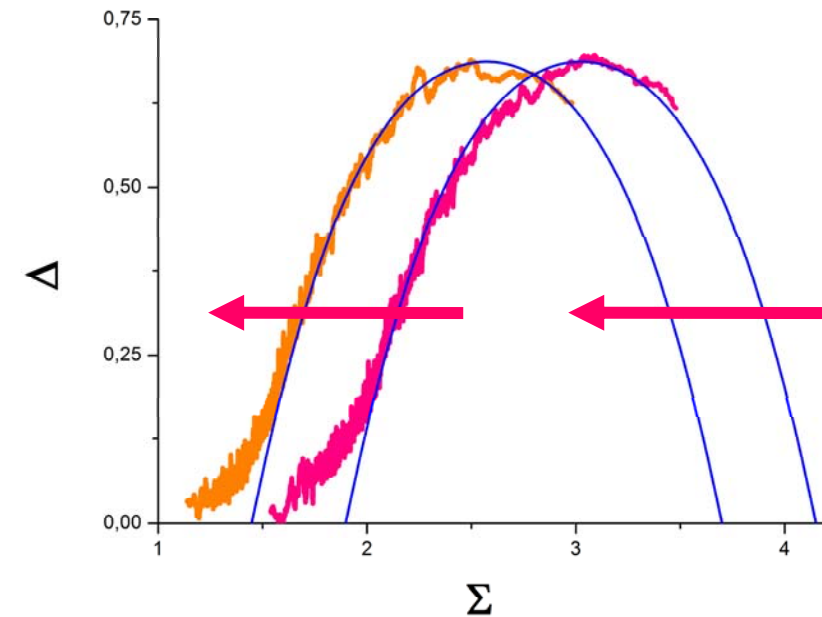
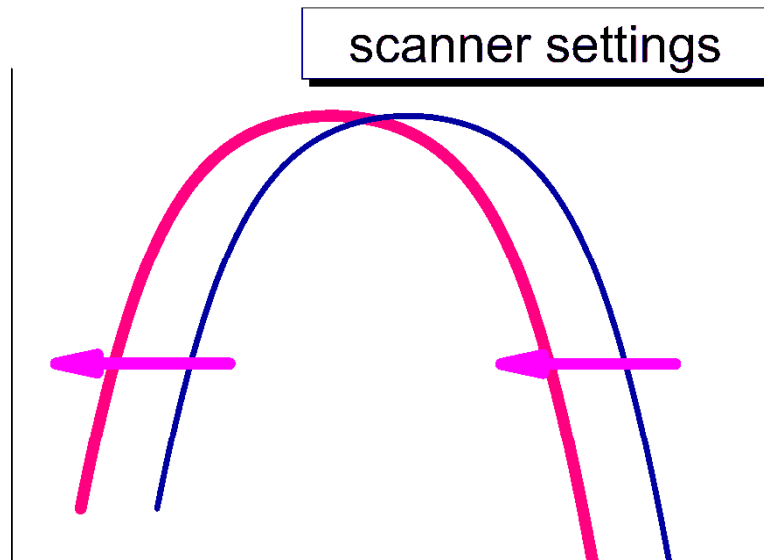
22,000

54,000

Improvements: probe selection:
genomic, thermodynamic
criteria

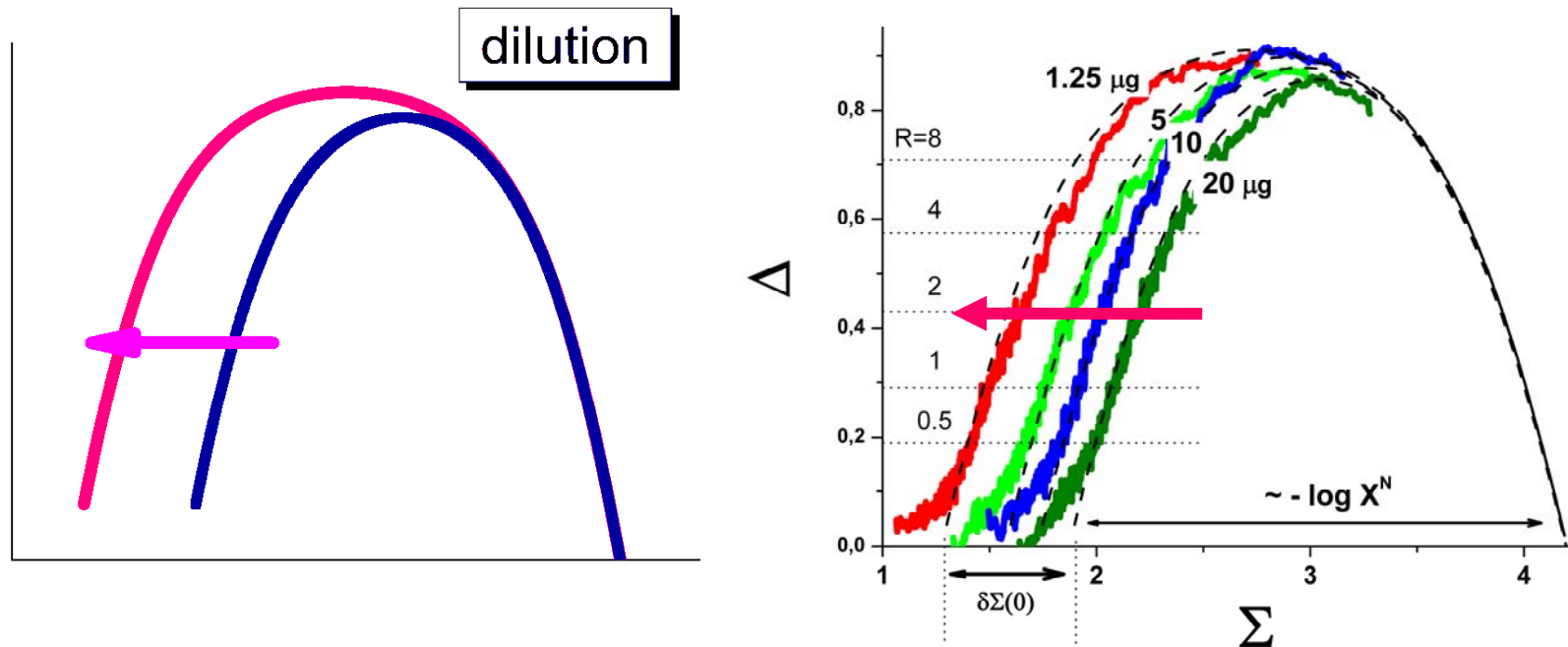
hybridization, labeling
kits, scanner etc.

Hook-pattern I: shift



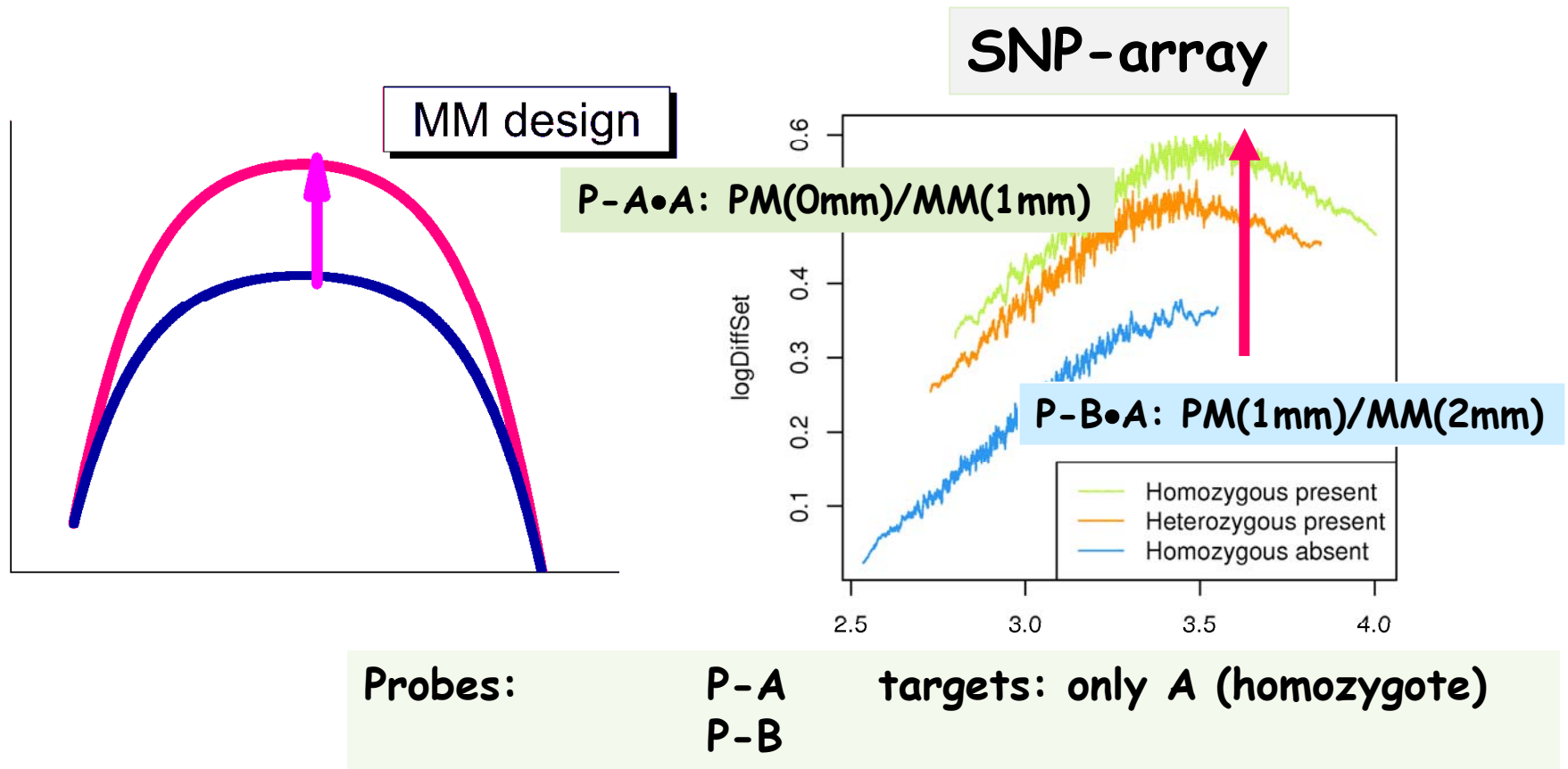
changes of the sensitivity-settings
(scanner, amount of labelling) shift the
whole curve in horizontal direction

Hook pattern II: widening



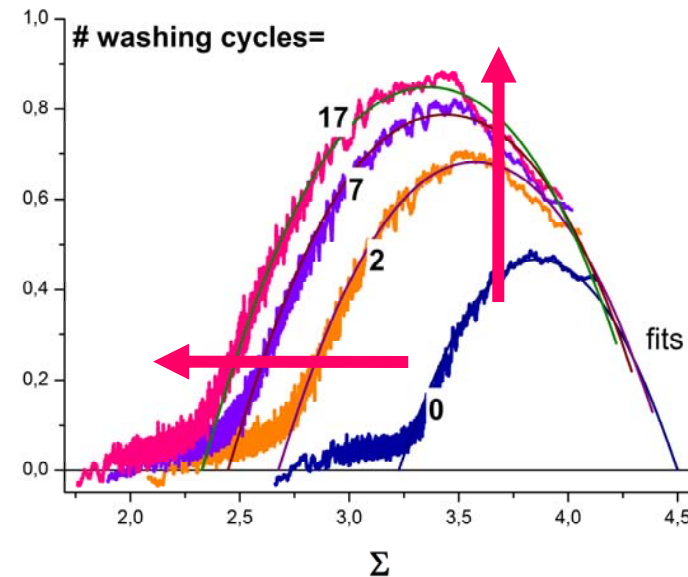
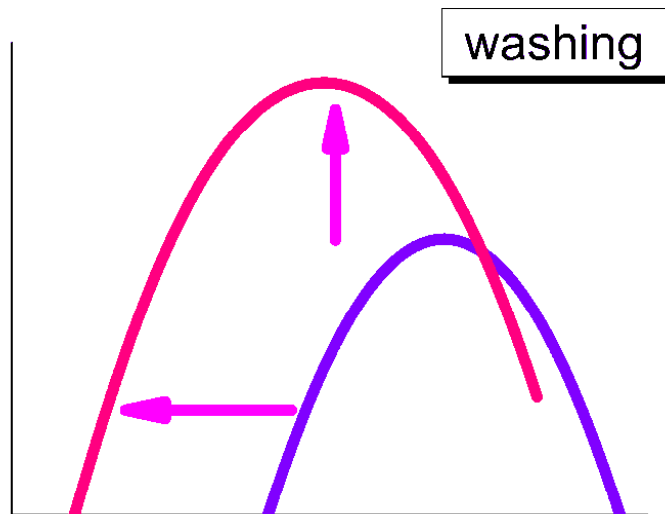
dilution (less total RNA) increases the width of the hook

Hook pattern III: vertical stretch



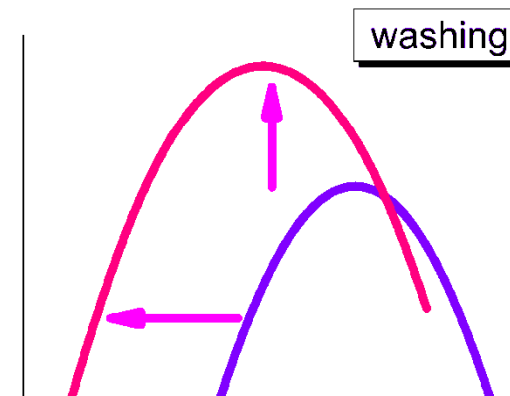
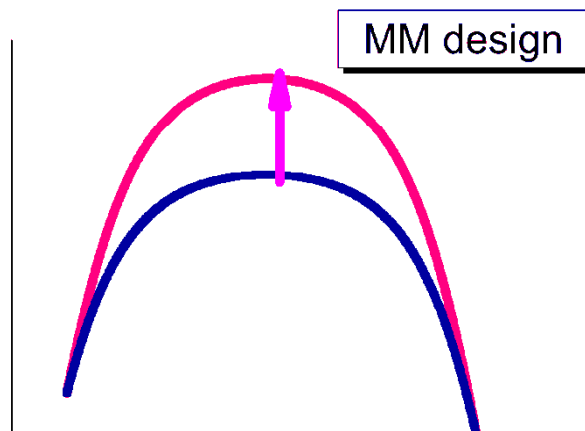
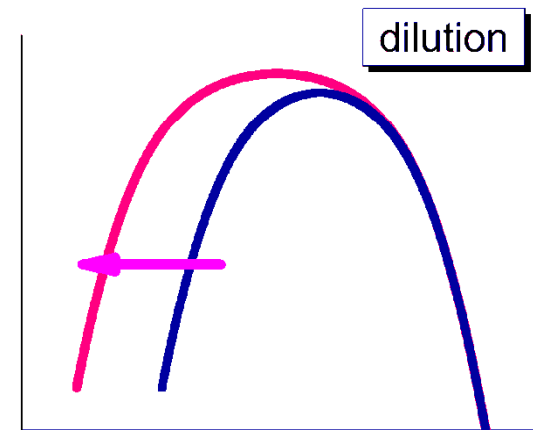
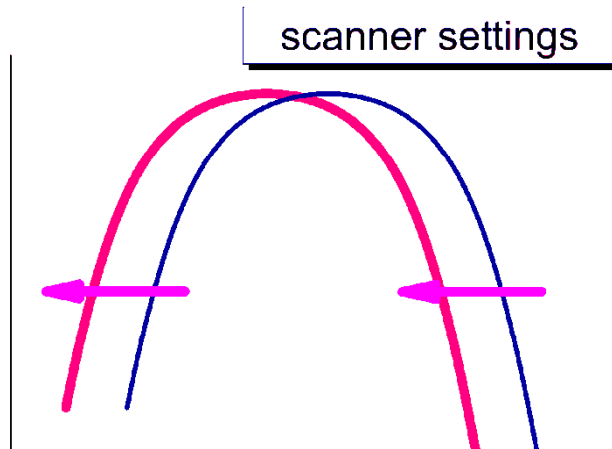
PM/MM-gain modifies the height of the hook

Hook pattern IV: „blow up“

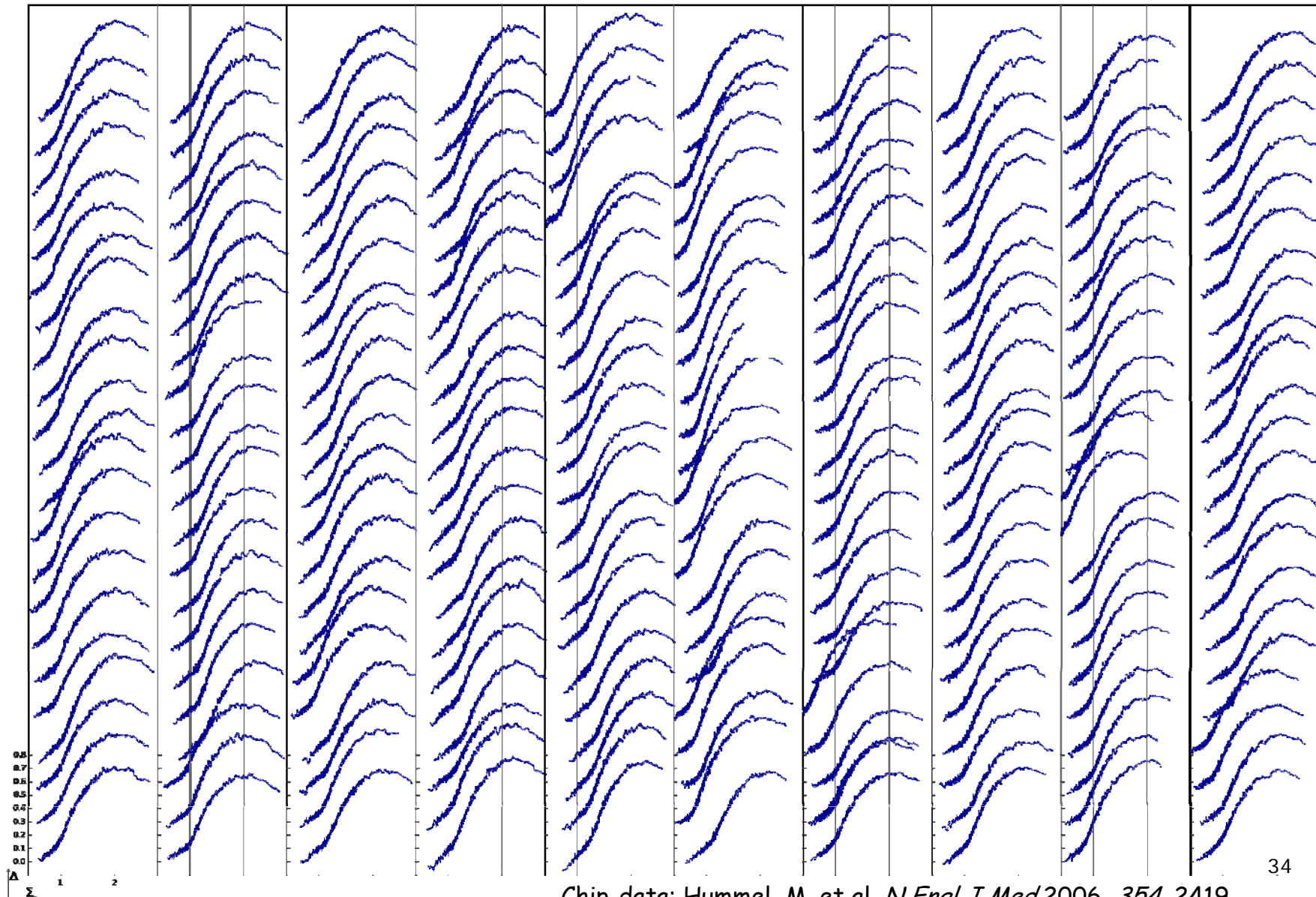


washing increases the width AND the height of the hook

Simple rules reveal underlying effects...

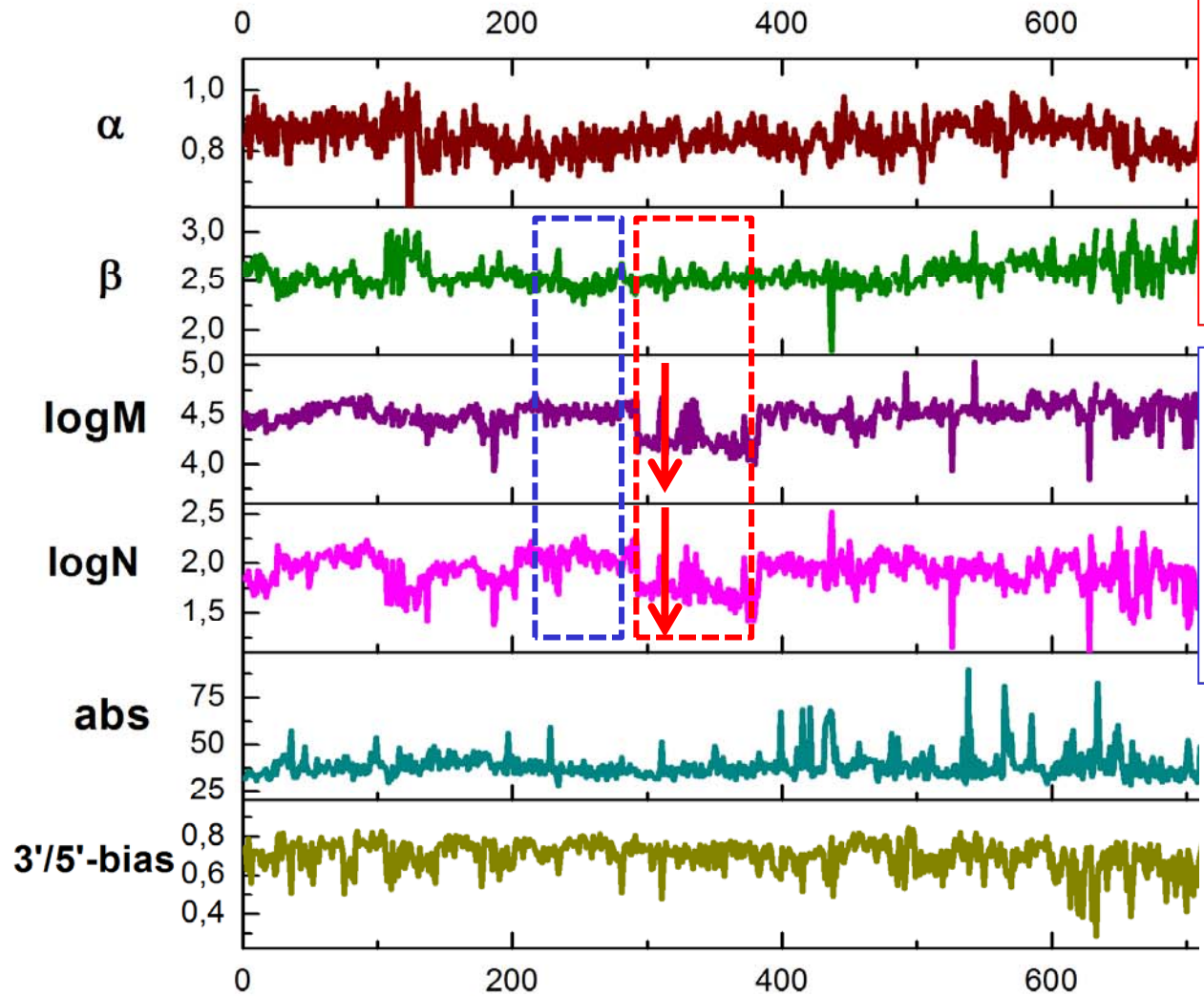


Clinical trial: Burkitts Lymphoma: 221 patients

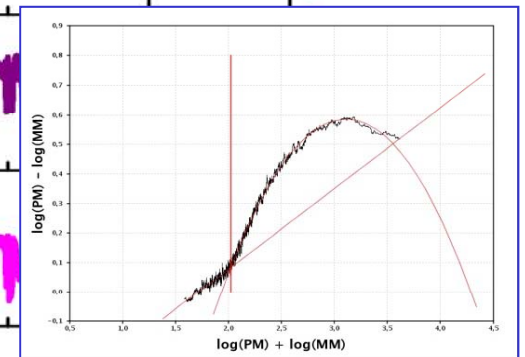
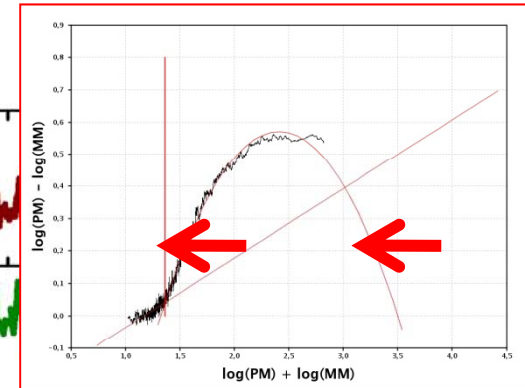


Chip-data: Hummel, M. et al. *N Engl J Med* 2006, 354, 2419.

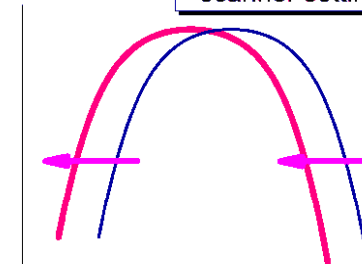
Scanner sensitivity



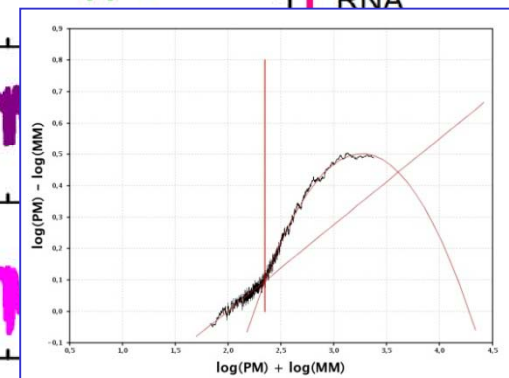
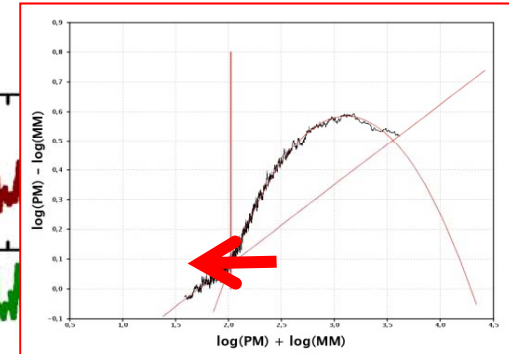
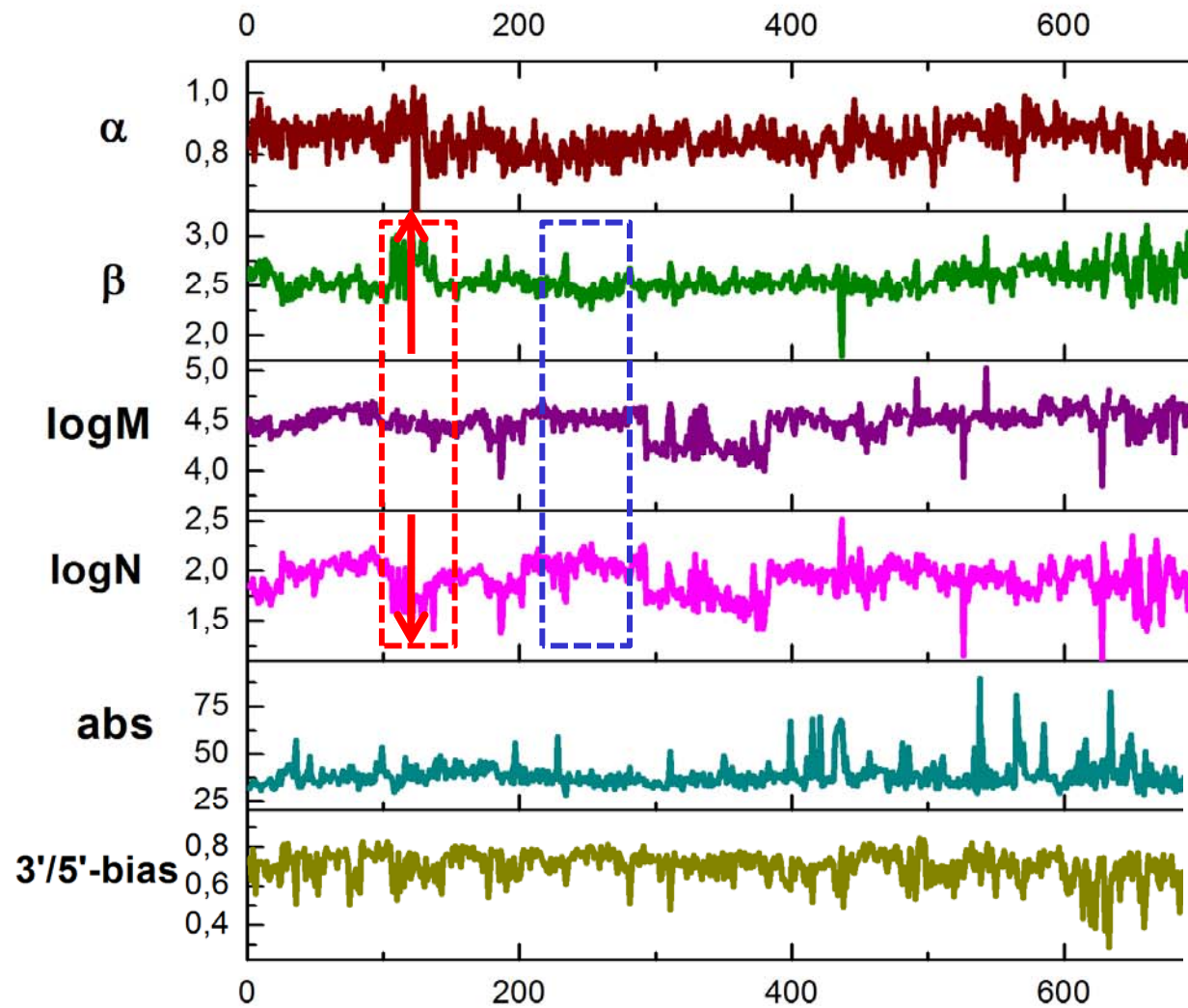
probe # ~ time (3 years), laboratories, samples



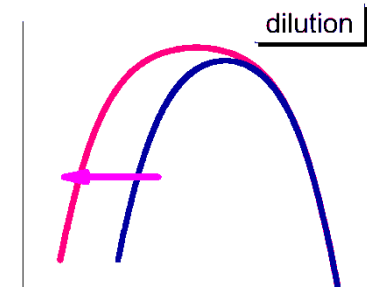
more
scanner settings



Dilution (less RNA (30%))

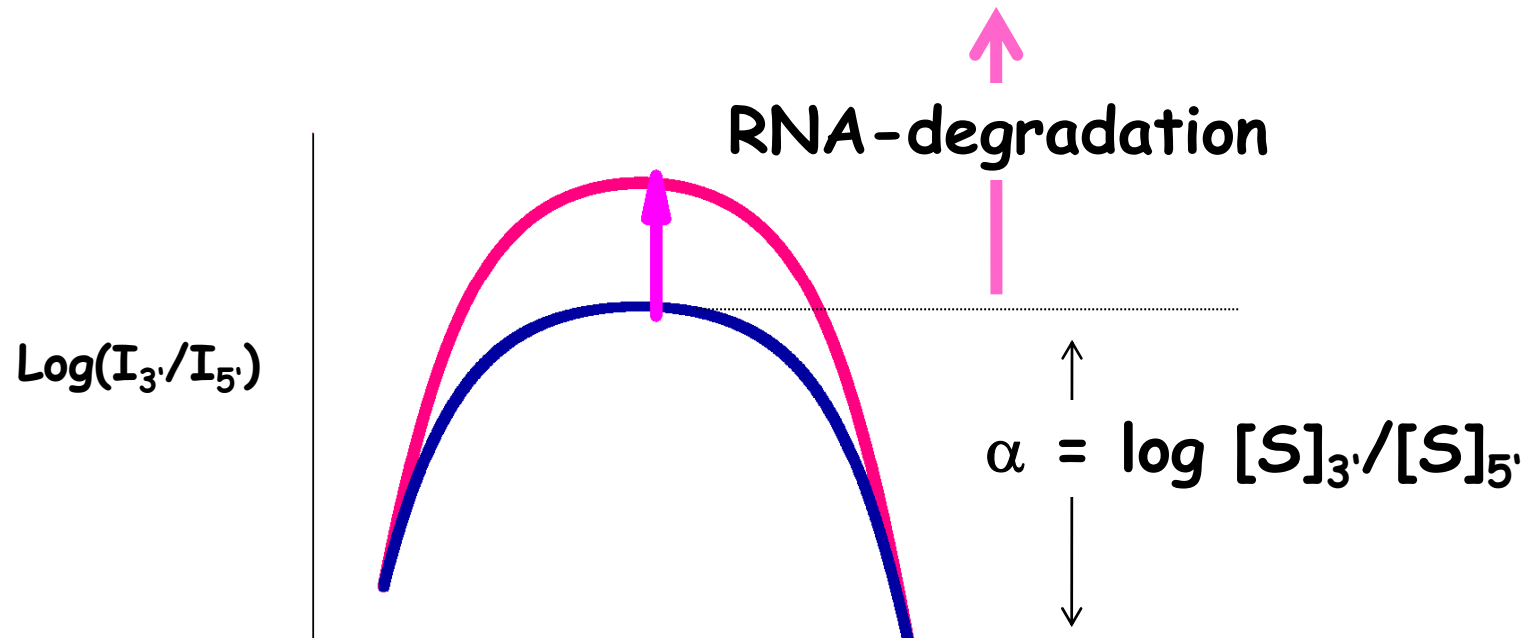


more



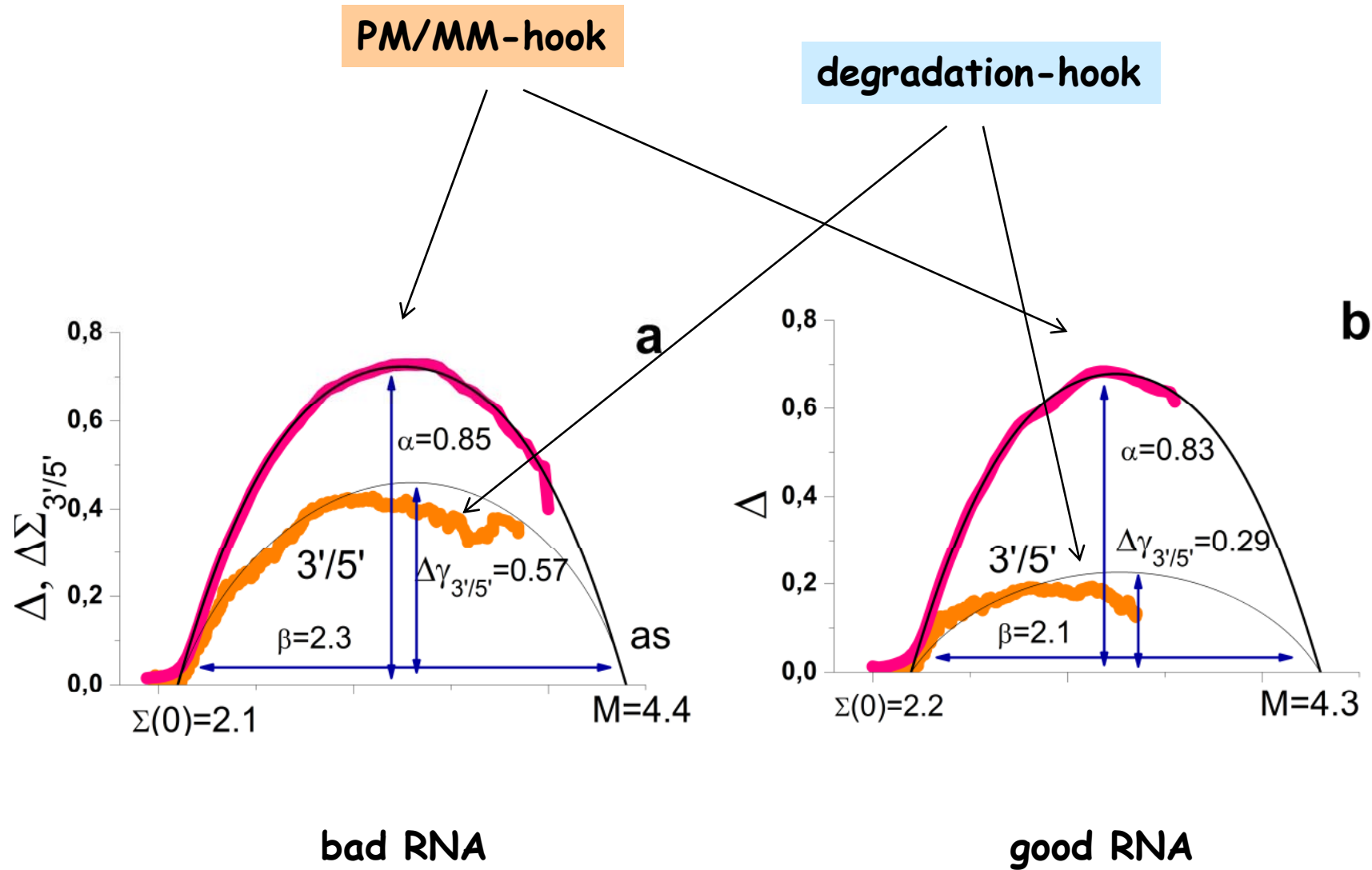
probe # ~ time (3 years), laboratories, sampl

degradation hook



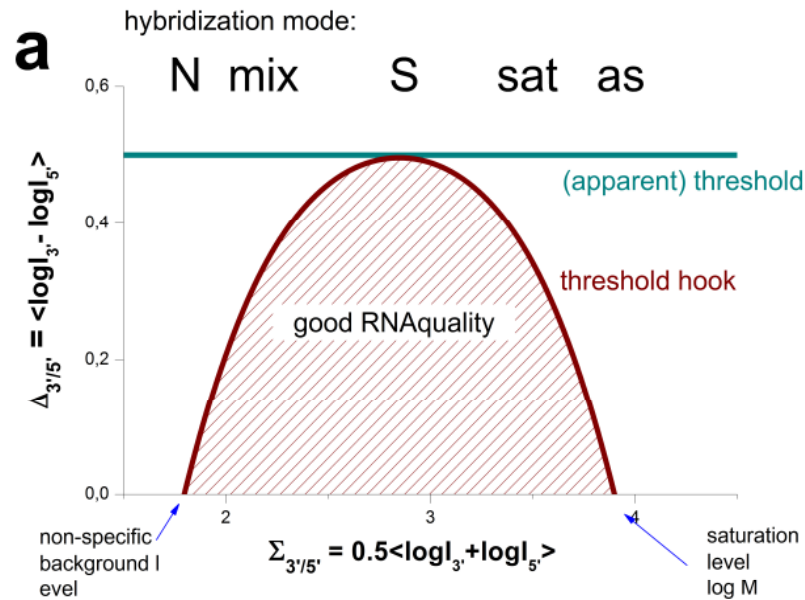
Depletion of targets with increasing distance from 3'-end defines the height of the degradation hook

degradation hook: example

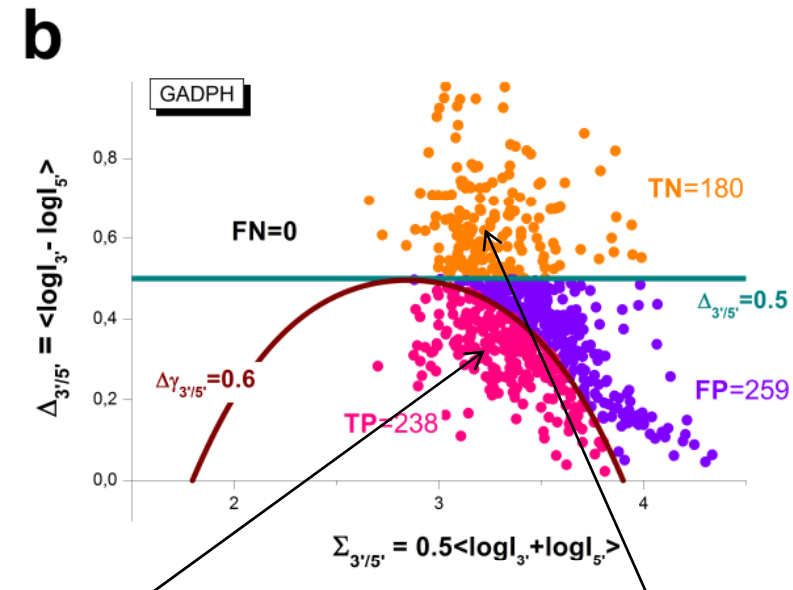


Degradation threshold

Threshold hook



Human tissue data



$$\text{Positive Predictive Value} = \frac{TP}{TP + FP} = 0.5$$

$$\text{Specificity} = \frac{TN}{FP + TN} = 0.8$$

Thank you !

Papers see

<http://www.izbi.uni-leipzig.de/izbi/mitarbeiter/Binder/binderpubl.php>